TRANSTHYRETIN (PREALBUMIN) IN HEALTH AND DISEASE: Nutritional Implications

Yves Ingenbleek

Department of Food Sciences, University Louis-Pasteur, 67401 Illhirch Cedex, Strasbourg, France

Vernon Young

Laboratory of Human Nutrition, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139

KEY WORDS: conformation, metabolism, nutritional status, amyloidosis, acute and chronic stressful conditions

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INTRODUCTION

Transthyretin (TTR), formerly called prealbumin, is a plasma protein secreted mainly by the liver and involved in the transport of both thyroid hormones. TTR is also indirectly implicated in the carriage of vitamin A through the

mediation of the retinol-binding protein (RBP). A number of recent reviews encompass all behavioral aspects of RBP in health and disease (27, 99, 217, 227). To our knowledge, no such information exists concerning TTR, with the notable exception of one survey published in 1987 in French (41). The present article aims to fill a gap in the Anglo-Saxon literature and to provide within a historical perspective a general overview of much that is known about TTR. Hence, this paper is an attempt to collect the most fundamental and clinical data currently available. We focus more specific attention on nutritional aspects because TTR is regarded as one of the most common biochemical indicators of protein-depleted states both in developed and developing countries. This review marks the twentieth anniversary of the first publication of TTR as a marker of nutritional status (125).

ANALYTICAL PROCEDURES

TTR was first identified in 1942 in concentrated cerebrospinal fluid (CSF) using the Tiselius moving boundary electrophoresis technique (148) and later by immunoelectrophoresis (88). The presence of TTR was soon confirmed in human plasma using several electrophoretic systems based on varying migrating media, such as paper sheet (278), starch gel (274), polyacrylamide gel (241), or reverse-flow electrophoresis with several buffering systems (122, 231). At the same time, pioneer groups working in the field of thyroid studies devised an alternative approach that assessed the maximal binding capacity of TTR for radiothyroxine (124, 207). The principle of the method was based on the addition of increasing amounts of labeled material in the incubation medium until the saturation of TTR binding sites was achieved. Under normal conditions, the plateau level recorded by maximal radiohormonal binding determines the TTR concentration of the unknown blood sample. More recent procedures measure TTR with affinity chromatography by the mediation of RBP coupled with sepharose (163) or by reversed-phase high-performance liquid chromatography (36).

The isolation of human TTR in a purified form (63, 224) and the preparation of specific antibodies gave impetus to the development of a large spectrum of immunological methods. Radial immunodiffusion (275), the oldest and simplest immune method, is still the most widely employed, followed by electroimmunoassay (164), immunofluorescence (95), turbidimetry (48), nephelometry (96), enzyme-linked immunosorbent assay (152), and radioimmunoassay (56). The recent advent of monoclonal antibodies (51) has boosted immunochemical studies in living tissues, making it possible to identify molecular variants of TTR.

The measurement of TTR either by its maximal binding capacity for radiothyroxine or by an immunological technique yields highly positively correlated results (129). However, this observation is valid only when the blood concentration of thyroxine-binding globulin (TBG), the most important of the three carrier proteins of thyroid hormones, remains within its normal dispersion range. The specific values are determined by gender and ethnical factors. The former approach becomes clearly irrelevant in the case of subnormal TBG levels, as seen in protein malnutrition (126). Under these circumstances, a spuriously high peak of radioactive thyroxine is displaced toward and accumulates in the prealbuminic zone, thus leading to an overestimation of TTR values (128). From these divergent results, one may conclude that when a genetic or acquired TBG deficit is likely or suspected, only an immunological method will reliably estimate the true TTR concentrations.

The isolation and purification of rat TTR (28, 196) has paved the way for the development of a useful animal model designed to provide further insight into thyroid, retinoid, and nutritional investigations. However, results obtained through these experimental studies must be interpreted with caution because in contrast to the human situation, TTR is the main transport vehicle of thyroid hormones in the rat. TBG plays a relatively minor role, circulating in the bloodstream during a limited period of the animal's growth (247).

CONFORMATIONAL STUDIES OF TRANSTHYRETIN

TTR appears as a nonglycosylated tetrameric holoprotein displaying tetrahedral symmetry. It consists of 4 identical subunits whose primary sequence of 127 amino acids (AAs) has been established, and it has a molecular mass (MM) of 54,980 Daltons (Figure 1, 150). The secondary and tertiary structures of each monomer have been studied using X-ray diffraction analysis of the crystalized protein at a resolution of 1.8 Å (25). Approximately 55% of the AA residues in each monomer are engaged in the formation of two extended eight-pleated sheets, each of which comprises four strands with antiparallel orientation. Only 5% of the AA residues are situated in one segment of α -helix (25). Two subunits aggregate to form a dimer by the interaction of two internal planes of the β -pleated sheets. The assembly of two dimers occurs through noncovalent links between two interplaying sites that belong to the edge strands of the sheets and that involve three hydrogen bonds and five AA residues (25).

The quaternary structure of TTR has the shape of a globular protein whose overall size is 70 Å \times 55 Å \times 50 Å. The two dimers are slightly rotated in relation to each other along the y axis. This positioning gives the molecule outstanding resistance to most alkali and acid solutions as well as to usual denaturating agents such as urea, guanidine-HC1, and sodium-dodecyl-sulfate (30, 31). The four subunits delineate through the molecule an open channel whose internal wall is formed by inner β -sheets and where two binding sites for thyroid hormones are located (25, 201). These binding sites are interrelated

NH₂-Gly-Pro-Thr-Gly-Thr-Gly-Glu-Ser-Lys-Cys-Pro-Leu-Met-Val-Lys-Val-Leu-Asp-Ala-Val-Arg-Gly-Ser-Pro-Ala-Ile-Asn-Val-Ala-Val-His-Val-Phe-Arg-Lys-Ala-Ala-Asp-Asp-Thr-Trp-Glu-Pro-Phe-Ala-Ser-Gly-Lys-Thr-Ser-Glu-Ser-Gly-Glu-Leu-His-Gly-Leu-Thr-Thr-Glx-Glx-Gln-Phe-Val-Glu-Gly-Ile-Tyr-Lys-Val-Glu-Ile-Asp-Thr-Lys-Ser-Tyr-Trp-Lys-Ala-Leu-Gly-Ile-Ser-Pro-Phe-His-Glu-His-Ala-Glu-Val-Val-Phe-Thr-Ala-Asn-Asp-Ser-Gly-Pro-Arg-Arg-Tyr-Thr-Ile-Ala-Ala-Leu-Leu-Ser-Pro-Tyr-Ser-Tyr-Ser-Thr-Thr-Ala-Val-Val-Thr-Asn-Pro-Lys-Glu-COOH.

Figure 1 The AA sequence of each TTR monomeric subunit (150).

by the phenomenon of negative cooperativity (10, 78), which implies that when one binding site is occupied by a first thyroid molecule, the association constant (Ka) for the second thyroid molecule is markedly reduced. Under normal conditions, the Ka of the two TTR binding sites for thyroxine (T4) has been estimated at 7×10^7 M⁻¹ and 6.7×10^5 M⁻¹, and that for triiodothyronine (T3) around 1.4×10^7 M⁻¹ and 5.5×10^5 M⁻¹ (47, 233). The phenolic group of the thyroid molecule stands in front of the hydrophilic group of the TTR tetramer in the narrowest part of the channel, whereas the opposite alanyl moiety floats in its wide-mouthed opening (25).

The external surface of the TTR tetramer is depressed by the presence of two deep concavities whose centers are ~ 33 Å apart, or the same order of magnitude as that separating the two DNA helices (25). The observation of a three-dimensional building structure complementary to the DNA edifice prompted Blake & Oatley to postulate that the plasma TTR could be the remnant of a phylogenetically related nuclear binding protein, which could serve as a model for thyroid hormone receptor studies (26).

TTR possesses one binding site for RBP, the specific carrier protein for one molecule of all-trans-retinol (Figure 2, 149). RBP is a monomeric chain comprising 182 AA residues (226) and totaling 21,000 Daltons as MM (27, 99, 149, 217, 226, 227). The conformational structure of RBP and its binding site for retinol have been determined by X-ray crystallography (198). In most instances, RBP is transported in the bloodstream in the form of a saturated holo-RBP protein equimolarly attached to TTR, which means that the resulting trimolecular retinol circulating complex (RCC) has an overall MM of \sim 76,000 Daltons (27, 99, 149, 198, 217, 226, 227). The Ka for the TTR-RBP binding site has been estimated at $1.2 \times 10^6 \, \mathrm{M}^{-1}$ (296). The TTR-RBP binding domain is still not entirely identified but rather is considered stereochemically unrelated to the AA sequence responsible for the antigenic response of the TTR molecule. This antigenicity has been localized in a midregion fragment of the

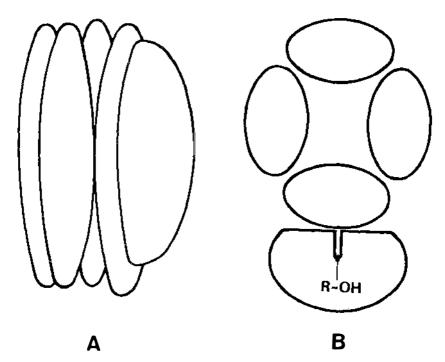


Figure 2 Schematic configuration of the retinol circulating complex (RCC).

- A. Lateral view showing the tetrameric structure of TTR (54,980 Daltons as MM). Under normal conditions, a single RBP molecule (21,000 Daltons as MM) is bound to one of the four subunits.
- B. Transversal view displaying the open channel delineated by the tetramer, which contains two binding sites for thyroid hormones. The inner part of the small RBP molecule has a cleft comprising one specific binding site for one retinol (R-OH) molecule.

monomeric subunit and corresponds to the 35-103 AA sequence, which forms at least two epitopes (51).

TTR has been identified in at least 15 vertebrate species (163). Most animal TTRs exhibit striking similarities to the human homolog, in particular to the tetrameric structure devoid of a carbohydrate load. The complete AA sequence of the TTR subunit has been clarified in three rodent species, namely rat (196), mouse (305), and rabbit (283), amounting to 127 residues like in human beings. Nevertheless, some minor interspecies differences in the AA sequencing hinder the cross-reaction between the specific animal antibodies and human TTR.

TRANSTHYRETIN GENE EXPRESSION

The nucleotide sequence of the entire human TTR gene, including the 5' (transcription initiating site) and the 3' (untranslated region) flanking regions,

has been determined (244, 290). The human gene has been localized in the long arm of chromosome 18 (306). The TTR subunit is encoded by a 6.9-kilobase (kb) gene composed of 4 exons and 3 introns (244, 290). Exon 1 contains 95 basepairs (bp) and 26 bp 5' untranslated and codes for a 20-AA leader peptide and AAs 1-3 of the mature protein. Exons 2 (131 bp), 3 (136 bp), and 4 (253 bp) possess the coding sequences for AAs 4-47, AAs 48-92 and AAs 93-127 of the monomer, respectively (244, 290). A common sequence upstream to the 5' cap site is highly conserved in different species and seems to play a critical role in gene expression (54). In addition, a liver-specific nuclear receptor has been discovered that can bind a distant enhancer element (55).

The TTR-mRNA spans ~ 0.7 kb and contains a 5'-untranslated region (26–27 nucleotides), a coding region (441 nucleotides), and a 3'-untranslated region (145–148 nucleotides) preceding the poly(A)tail (182, 272). Human (182, 306), rat (67, 283), and mouse (54, 305) coding regions exhibit a considerable degree of homology (~ 85%). This similarity suggests a phylogenetically preserved modulating role in gene expression. As predicted by the cDNA nucleotide sequence (182), TTR-mRNA encodes the pro-TTR-monomer (pre-TTR), a polypeptide comprising 147 AA residues whose N-terminal region is a hydrophobic signal peptide consisting of 20 AAs (142, 272). Pre-TTR undergoes a cleaving process during its migration through the endoplasmic reticulum bilayer to yield the native TTR-monomer after leakage of the signal peptide (142, 272). During the secretory process, four identical subunits agglomerate to build the mature tetrameric molecule (150).

To our knowledge, no ethnic differences are reported for TTR, in contrast to the genetically high and low TBG serum levels described in African (126) and Australian (311) populations, respectively. Some authors have reported that TTR may exhibit microheterogeneity (218). Elderly people may be subjected to amyloid organ deposits principally confined to the myocardial tissue (52). Normal TTR participates in the senescent process (314). The mechanisms whereby nonmutated TTR takes part in this systemic amyloidotic deposit are not yet clear. This involutional phenomenon is a common feature of increasing age. It is found in the heart of 25% of individuals over age 80 (314) and has also been documented for other molecules of biological importance, such as ferritin, fibrinogen, and immunoglobulin light chains.

It has been demonstrated in recent years that the monomeric subunits of TTR may undergo a number of mutations consisting of the substitution of a single residue at different positions of the normal AA sequence. Andrade et al first described the clinical anomaly (9). At least 38 different variants have been identified (Table 1), all of them generated by a single mutation point in the coding region of the TTR gene. Some mutant forms (Ser-6, Arg-102, and Thr-109) were fortuitously discovered because they remain asymptomatic and

apparently nonpathogenic (5, 80, 186). Most reported TTR mutations are associated with varying degrees of extracellular deposit of amyloid fibrils in different tissues. Familial amyloidosis covers a wide range of inherited diseases with autosomal dominant transmission. The variable penetrance explains the heterogeneity of clinical manifestations whose extreme poles are (a) prominent cardiomyopathy, as seen in patients with Thr-45 (243), Ile-50 (202), Ala-60 (307), Leu-68 (6), and Met-111 (203) mutations, and (b) the familial amyloidotic polyneuropathy (FAP) entity identified by Costa et al (53) and found in patients with Met-30 (242, 285), Ala-30 (146), Leu-33 (111), Pro-36 (145), Leu-64 (121), Tyr-77 (308), and Ser-84 (73) mutations. The TTR-Ile-122 (102) is the sole amyloidotic mutation characterized by recessive inheritance. All known carriers of this last allelic form belong to the same African-American racial group, a finding consistent with the concept of a de novo mutation arising from a single founder (140). This senile systemic amyloidosis constitutes a distinct entity whose clinical expression, mainly cardiac, does not develop before age 70 (102, 140).

Table 1 indicates that exon 1 has not been incriminated thus far as a site of a point mutation and that six different AAs situated at positions 33-Phe, 47-Gly, 49-Thr, 50-Ser, 58-Leu, and 84-Ile of the normal monomeric sequence may diverge from normalcy along two distinct substitutive pathways, whereas 30-Val may undergo three different replacing processes.

The vast majority of the reported mutations remain very rare entities. The most common molecular defect leads to the substitution of methionine for valine at position 30 of the TTR monomeric sequence (72, 242, 285). The anomaly stems from a guanine to adenine replacement in the first codon, which creates a cleavage site in exon 2. The variant reaches its highest prevalence in Portuguese siblings but is also found in Swedish, Japanese, and Mediterranean families (72, 242, 285). Clinical symptoms are quite similar within these various kindreds and are identified as Type I FAP. Heterozygous siblings biosynthesize both normal and mutated TTR in varying proportions. The inherited abnormality is expressed very early in development (200), which allows prenatal detection by chorionic villus sampling. Homozygous FAP is a rare condition that has nevertheless been reported in some kindreds (118).

From a functional point of view, TTR variants involved in amyloid disorders may exhibit normal, increased (80, 186), or reduced (229) Ka for thyroid hormones. Distortions in Ka values presumably depend on the degree of interdependence between the defective molecular domain and the hormonal binding sites. There is no effective form of therapy for the inherited mutation, which implies that genetic counseling of the asymptomatic carriers is mandatory in exposed families. Liver transplantation in FAP patients is accompanied by clinical improvement (119), but the long-term outcome of this substitutive surgery remains to be evaluated.

Table 1 Transthyretin mutated variants described in the literature

Location and nature				
Encoding exons	of AA substitution	Ethnic origin	Reference	
Exon 2 (44 AAs: 47	6: Gly ── Ser	Nonamyloidotic	80	
	10: Cys → Arg	Hungarian	291	
	→Met	FAP I (Portuguese, Mediterra- nean, Swedish, Japanese)	72, 242, 2	
	30; Val → Ala	German	146	
	Leu	Japanese	195	
	Ile	Polish-Jewish	194	
	33: Phe <			
	Leu	Polish-Lithuanian	111	
	36: Ala → Pro	Greek	145	
	42: Glu → Gly	Japanese	293	
	45: Ala ──Thr	Irish-Italian	243	
	Arg	Japanese	191	
	47: Gly (
	Ala	Italian	79	
Exon 3 (45 AAs: 92	Gly	Jewish	220	
	49: Thr		_	
	Ala	Italian	7	
	Arg	Japanese	293	
	50: Ser	_		
	Ile	Japanese	202	
	55: Leu → Pro	Dutch-German	141	
	His	Maryland-German	199	
	58: Leu			
	Arg	Japanese	240	
	60: Thr → Ala	Appalachian	307	
	61: Glu → Lys	Japanese	258	
	64: Phe → Leu	Italian	121	
	68: Ile → Leu	German	6	
	69: Tyr ── His	Scottish-English	326	
	70: Lys — → Asn	German	138	
	71: Val → Ala	French	18	
	77: Ser → T yr	Illinois-German	308	
	Ser	FAP II (Indiana-Swiss)	73	
	84: Ile	Italian	264	
	Asn	Sicilian	204 7	
	89: Glu → Gln 90: His → Asn		262	
E 4 (25 4 4 127 02)		Italian Nonamyloidotic	5	
Exon 4 (35 AAs: 127 — 93)	102: Pro → Arg 109: Ala → Thr	Nonamyloidotic Nonamyloidotic	186	
		Danish	203	
	111: Leu → Met 114: Tyr → Cys		203	
	114: Tyr	Japanese French-Canadian	292	
	116: Tyr	German-Scandinavian	113	
	119: 1111 → Met 122: Val → Ile	African-American senile systemic	102	
	122. v ai ——Ile	Antean-American senire systemic	102	

PHYSIOLOGY AND METABOLISM

When the TTR molecule was first identified in the CSF (88, 148), its physiological function was unknown and was thus referred to as "component X" (88). Subsequent studies confirmed the presence of component X in both CSF and serum (278). At the time, the protein was characterized solely by a fast electrophoretic migration rate. Located just in front of the albumin band, TTR was the most anodal of all CSF and serum-moving proteins (88, 278). Soon thereafter, Schultze et al isolated component X from human serum (250). Its purification and chemical analysis led the German authors to recognize one of the salient characteristics of the protein, i.e. its relatively high richness in tryptophan. Hence the name "Trp-rich prealbumin," proposed by this working group (250).

The functional properties of TTR were partially disclosed in 1958 when Ingbar demonstrated its transport capacity for thyroxine (123), thus justifying the name of "thyroxine-binding prealbumin" (TBPA) given at that time. However, the appellation appeared somewhat restrictive when it became obvious that TBPA also conveyed triiodothyronine (60). Its inadequacy was further reinforced after the demonstration that TBPA was the unique vehicle bound to RBP within the RCC edifice (149). The name "transthyretin" given to the protein by the International Nomenclature Committee (98) emphasizes TTR's concomitant involvement in the transport of both thyroid hormones and of retinol. All previous names registered over four decades had pinpointed electrophoretic or chemical peculiarities of the molecule. As a result of advancing knowledge, the now universally accepted term "transthyretin" stresses the dual physiological roles played by TTR in vertebrate species.

Distinct secretory mechanisms govern the liver production of both TTR and holo-RBP. In particular, the hepatic release of holo-RBP (but not its synthesis) critically depends on the retinoid status (270). Pulse-chase studies using ³⁵Slabeled methionine incubated with isolated hepatocytes have shown that both carrier proteins are first identified in the endoplasmic reticulum and later transported to the Golgi complex prior to extracellular delivery (85). Like pre-TTR (142, 272), native RBP is processed through a precursor form of larger MM (273). The maturation of this pre-RBP is similarly characterized by the cleaving release of the signal peptide, which yields the physiologically active monopeptide as an end product (273). Both TTR and holo-RBP strongly coalesce in the bloodstream in a 1:1:1 equimolar ratio. This process allows the low MM holo-RBP to stabilize and prevents its rapid renal or intestinal leakage. The biological half-life of TTR is about two days (271), whereas that of holo-RBP within the RCC fluctuates and is about half a day (216). After the uptake of retinol by peripheral tissues, the resulting apo-RBP devoid of its ligand has a significantly shortened half-life of ~ 3.5 h (217). Immunofluorescent studies of renal tissues have confirmed the target role of the kidneys in RBP catabolism (95). In contrast to TTR, whose labeled molecules remain confined within the glomerular tufts, RBP diffuses rapidly in the tubular lumen and in intercellular spaces (95). Under normal circumstances, only minute amounts of RBP are recovered in the urinary output, which indicates that most molecules undergo tubular disintegration with subsequent recycling of their AA residues. Using animal models and radioiodinated TTR, Makover et al showed that the main site for TTR degradation is the liver, followed by muscle mass, skin, and kidneys (170).

TTR can be detected in fetal blood as early as eight weeks after conception (11). Fetal TTR likely originates from dual sources. One part seems to be inherited passively from the pregnant mother through transplacental filtration (94). The remainder results from fetal hepatic synthesis, as indicated by the presence of significant amounts of TTR-mRNA in the early developmental stages of this organ (171). The respective importance of these two TTR sources during fetal life remains to be clarified. Several groups have found diminished TTR levels in the umbilical cord of preterm neonates compared with healthy full-term newborns (23, 139, 245, 294). Increasing gestational age is accompanied by a slow and predictable rise of TTR values, which are correlated with birth weight and which proved useful in distinguishing between small, appropriate, and large-for-gestational-age infants (23, 245). The plasma concentration of TTR is age dependent, and in healthy neonates it is approximately half that found in adults (32, 129, 136, 236, 275, 294). During the entire prepubertal period, TTR increases progressively but with no differences in levels between the sexes (129, 236, 255, 275, 294). TTR increases sharply at the onset of puberty, with a more pronounced elevation in male than in female adolescents (130). The appearance of this sexual difference is determined by sex steroid hormones and maintained in the form of plateau levels during full sexual maturity (32, 130, 275), reaching 323 \pm 49 mg/liter (SD) in males and 283 \pm 43 mg/L (SD) in females (130). Plasma TTR concentrations begin to decline in both sexes after age 50, but along a steeper slope in elderly males than in females (130), so that a sexual difference no longer persists after the seventh decade of life. In serum from normal newborns, children, adults, and elderly persons, TTR and RBP interact with retinol in a close equimolar stoichiometry over all concentration ranges (23, 27, 99, 129, 130, 139, 217, 227, 270, 294). This interaction implies that the bulk (90-95%) of retinol in human blood circulates within the trimolecular RCC and that both apo-RBP and holo-RBP unbound to TTR represent only minor fractions.

It has been suggested that the sharp elevation of RCC at puberty could reflect an augmented demand for retinoids by peripheral tissues (227). The aforementioned influence of sex steroid hormones on TTR synthesis and/or turnover rate has been confirmed in several clinical conditions using natural or synthetic analogs (33, 241) as well as in women taking oral contraceptives with varying proportions of estrogens and progestagens (183). The reported hepatic production of TTR has been depressed by zinc deficiency (15) and stimulated by glucocorticoids (93, 208) but remained unaffected by vitamin A deprivation (197). Several research groups have explored the behavior of TTR in intestinal, hepatic, renal, and thyroid disorders (42, 122, 267, 268, 295). All of these physiological or diseased conditions may eventually modify either the blood concentration or the equimolar ratio between the three components of the RCC. For example, this is the case in normal pregnancy characterized by a relative increase of free holo-RBP (265), which may fulfill increased retinoid requirements. In contrast, free apo-RBP is present in molar excess in kidney patients (268) and in premature infants (253), probably as the result of impaired catabolism, reduced excretion, (268) or primary lack of intrahepatic retinol reserves (212). The multifaceted functional consequences of these alterations await elucidation.

In addition to hepatic production, three other TTR synthesis sites have been identified in mammals: visceral yolk sac endoderm (266), retinal pigment epithelium (172), and choroid plexus epithelium (116). Because the bloodbrain barrier is not freely permeable to thyroid hormones, for a long time investigators suspected that a locally secreted protein served as its intermediate carrier to the CSF. In situ TTR production within the central nervous system (CNS) was demonstrated by intense immunochemical staining of the organelles involved in the CSF secretory processes (3) and by the in vitro culture of human fetal choroid plexus (142). Animal experiments have shown substantial amounts of TTR-mRNA in specific CNS regions, ranging from 11 (272) to 30% (249) of the hepatic level. The highest TTR-mRNA concentration accumulates in the epithelial cells lining the ventricular surface of the choroid plexus (277) and seems to account for most, if not all, of the TTR found in the CSF (105). Although both TTR and albumin reveal very low CSF levels (~0.017 g/liter and 0.02 g/liter, respectively), their intrathecal ratio is ~30-fold higher than that found in plasma (313), which precludes their passive diffusion from the bloodstream. TTR levels remain stable in CSF between the ventricular and lumbar regions, in contrast to the increasing concentration gradients observed for albumin and immunoglobulins G (313). Considering the weight difference between liver and choroid plexus as well as the difference in the TTR concentration between plasma and CSF, it is assumed that the synthetic rate of TTR by the choroid plexus would need to be 13 times faster (249) to reach its usual intrathecal values, which represent ~ 20% of total CSF proteins (313).

The production of TTR in the liver and in the choroid plexus is regulated independently, notably during malnutrition and inflammatory processes (68, 304). Recent studies have proposed a transport model whereby TTR could be

involved in the uptake of thyroxine from the bloodstream and its delivery into the CSF (249). TTR thus appears to be a major hormonal carrier protein, conveying up to 80% of the intrathecal thyroxine (105). These data strongly suggest that TTR fulfills important ontogenic and functional properties in mammalian nervous structures, a concept further corroborated by the observation of its increased concentration in CSF during the neonatal period (162). The recent demonstration of intraocular RBP synthesis (173) makes it likely that local TTR (172) cooperates in the transport of retinol and in the visual cycle. The reduced solubility of the TTR variants could explain the frequent ocular damage in patients suffering from amyloidotic disorders (264).

At least two other potential physiological properties have been attributed to TTR. The first is related to the discovery of a binding site with a high Ka for epinephrine (65). The other is based on the description of a persisting thymic hormonal-like activity of thymectomized mice (37). This functional effect seems to be intrinsic to the TTR molecule and has been assigned to the first 10 AA residues of the TTR monomeric sequence (38). Thymulin is a zinc-dependent nonapeptide involved in the maturation and replication of the thymic T-cell lines (12), whose production patterns are markedly disturbed in protein-depleted states (46, 302). Finally, TTR has a sequential homology with several gastrointestinal hormones of the glucagon-secretin family (147), whose phylogenetic and biological significance remains unclear. Nevertheless, these molecular approaches raise new questions about the relationships between alterations of nutritional status, immune responses, regulatory peptides, and biogenic amines.

TRANSTHYRETIN IN NUTRITIONAL SURVEYS

Significant alterations in the levels of protein and calorie intakes by animals and humans affect protein turnover, synthesis, and breakdown. This situation entails a number of adaptive mechanisms in the body's N economy. Specifically, the visceral compartment reacts quickly to alterations in nutrient supply. Hence, clinicians have drawn attention to several visceral markers, such as serum-albumin (SA), transferrin (Tf), TTR, RBP, and transcortin (corticosteroid-binding globulin, CBG), which are regarded as potential indicators of this compartment. This conclusion relies on the assumption that, in uncomplicated fasting or starvation, fluctuations of the visceral protein compartment precede and reflect an overall but slower change in the total body protein.

The first proposal using TTR as an index of protein and energy malnutrition (PEM) appeared in *Lancet* 20 years ago (125). The study was performed on 40 PEM children hospitalized in Dakar (Senegal, West Africa) using radial immunodiffusion and specific anti-TTR antibodies. The micromethod only required a few microliters of serum drawn from the fingertip at weekly intervals

throughout the refeeding period. The proposal of TTR as a nutritional index was based on the observation that the drop of both TTR and SA demonstrated comparable restoration during dietary protein replenishment. Nevertheless, the rates of recovery of both biochemical indices revealed striking differences, showing that TTR was characterized by a faster response (125, 127). The persistence of a TTR:RBP:retinol molar ratio along all steps of nutritional depletion and recovery despite significantly different biological half-lives led Ingenbleek et al to propose that the level of TTR synthesis by the liver was the primary factor responsible for the peripheral retinol status (127). This view was recently confirmed under experimental conditions using a mutant mice model carrying a disruption at the TTR locus (74). This finding may also explain why overlooking the crucial role of TTR in vitamin A metabolism resulted in unachieved or even misleading conclusions.

The peculiar potential of TTR to detect subclinical PEM was attributed to the conjunction of at least three factors: its production by the liver and early response to nutritional deficits, its short biological half-life, and its unusual richness in the indispensable Trp AA (IAA) (125, 127). The first two points are broadly accepted, but the last one remains to be validated. Substantial advances in knowledge have been recorded over the last two decades in relation to protein and energy metabolism under conditions of health and uncomplicated malnutrition. Here we pinpoint some salient features to pave the way for a deeper scrutiny of more complex situations, especially those in which nutritional and inflammatory stresses coexist. These approaches constitute the prerequisite for a more objective assessment of nutritional status and nutrient requirements, taking into account the role of sex, age, and varying physiological and diseased states.

Protein synthesis is usually considered to start with the transcription of DNA into mRNA. The production of protein molecules is achieved by the ribosomal machinery, using 20 AA residues as building blocks. Several nutritional variables, working in concert or independently, may modulate the operative mechanisms at a translational (276), transcriptional (193), or posttranscriptional (320) level. Most energy-dependent processes (synthesis, folding, secretion, transport, and breakdown) associated with protein turnover and culminating in functional properties have been clarified progressively over the last few years (81, 323, 325). The sensitivity of body N to changes in energy intake has been recognized for many decades (40, 188), and this topic has been the subject of recent reviews (76, 322, 325). Nevertheless, there is a continuing uncertainty about the most effective sources and levels of energy-yielding substrates and the proportion among these for the support of protein anabolism. Although it is beyond the scope of this review to examine these aspects in detail, we must note that glucose functions as a major substrate under a wide range of pathophysiological conditions (246). If carbohydrate intake is restricted, glucose must be synthesized by gluconeogenesis, primarily from the AAs present in endogenous and dietary proteins. The direct provision of fatty acids alone appears ineffective in stimulating protein synthesis in rat hepatocytes (320) or in improving N sparing in fasting patients (24, 188). The beneficial effects of triglycerides as an energy substrate source on protein synthesis operate through the energy released by the β -oxidation of free fatty acids (used in part to promote glucose production from lactate within the Cori cycle) (39) or by the hepatic conversion of their glycerol moiety into glucose (34).

The relative proportion of dietary AAs and energy-yielding substrates that most effectively supports protein synthesis is currently a field of active investigation, and TTR is clearly at the cutting edge of the controversy. Rat experiments have established that both protein and energy restriction inhibit the generation of cytosolic TTR-mRNA (61, 165), thus resulting in the expected drop of plasma TTR to below control values (77, 303). The current opinion is that the rate of hepatic protein synthesis is critically dependent on appropriate energy provision and correlated to the bioavailability of individual free amino acids (87). Clinical surveys in fasting obese patients led some investigators to believe that energy supply rather than protein intake was the major determinant of visceral protein synthesis (256), whereas others have concluded that dietary protein is of greater significance when studies last for as long as 98 days (35). These apparent contradictions can be reconciled if TTR is an indicator of the protein metabolic status of the visceral compartment, as discussed in greater detail below. This concept is supported by the observation of Moscowitz et al in preterm infants given varying dietary energy:protein ratios (185). They concluded that although the AA supply is the main factor for maintaining protein synthesis, it can be affected by fluctuations of energy intake (153). Furthermore, this concept corroborates previous data showing that the visceral protein synthetic rate may be maintained at a higher level by glucose-free AA than by N-free dextrose preparations (263) or that the N balance may be preserved despite severely restricted energy dieting (214, 323). The former condition necessitates a higher-than-normal proportion of dietary AAs, which undergo deamination and gluconeogenic conversion to replace the lacking fuel. This increased AA oxidation corresponds to a form of nutritional wastage, which augments the N cost of protein synthesis, as documented by an increased urinary excretion of urea.

The protein-sparing effect of AAs in the absence of an adequate energy supply depends on the disposal of AAs themselves (103) and is unrelated to changes in insulin (84) or in fat mobilization (239). Piecing these elements together, declining TTR plasma levels appear to be the direct consequence either of an insufficient and/or inappropriate AA regimen or of a metabolic state that alters the balance between protein synthesis and degradation. The

former explanation seems likely for phenylketonuric children (254) and the latter for diabetic patients (89) with an inadequate temporal pattern of therapy. Under such circumstances, the decline in TTR plasma values may be blunted to some extent or, on the contrary, further depressed by increases and/or decreases in dietary energy. These modulating effects generated by glucose on TTR production are assumed to be dependent on its ability to redress a disrupted N/energy balance, which allows the recovery of a proportion of IAAs that would otherwise be degraded via gluconeogenic processes. Here we recall the pioneering studies, notably those of Lavoisier, Prout, Chevreul, and Liebig, initiated at the turn of the nineteenth century, which led to the identification of the major nutrient classes and of the main chemical constituents of carbohydrates, fats, and proteins. Shortly thereafter, Claude Bernard became one of the first investigators to establish a link between nutrient categories by ascertaining that "lipids are burning in the flame of sugars" (19). We consider that the measurement of rapidly turning over proteins for nutritional evaluation purposes expands upon the earlier concept by also asserting that "the synthesis of proteins occurs in the flame of sugars."

In this metabolic context, the potential role of Trp is of peculiar interest. A number of basic considerations are worth pointing out. First, Trp accounts for the lowest concentration of all IAAs in usual foodstuffs and in mammalian tissues (189). Second, it was demonstrated early in this century that adult rats given a Trp-deficient diet rapidly lost weight at a rate similar to that observed with a protein-free regimen (211). Third, Trp can reaggregate polyribosomal subunits of protein-depleted cells, thereby restoring the protein-synthesizing machinery (260). Fourth, as mentioned above, the AA sequence of both TTR and RBP is characterized by an unusually high Trp content (150, 226). Hence, it is tempting to speculate that the production of these Trp-rich carrier proteins (125, 127) is particularly sensitive to food protein restriction. The concept that Trp shortage might serve as a limiting factor for TTR synthesis was recently tested in vivo in growing rats submitted to acute and severe Trp depletion and compared with pair-fed and control groups (29). This experiment failed to confirm the foregoing hypothesis, but the conclusion drawn by these investigators may not be valid because the three animal groups exhibited divergent growth rates. Consistent with previous findings (211), the Trp-deficient group alone underwent dramatic food intake and body weight reductions, which implies substantial tissue (mainly muscle mass) depletion. Because ~ 80% of the AAs released by protein breakdown may be recycled (322), observed data (29) might well compensate for the reduced Trp intake so as to maintain unaltered protein synthesis rates. Obviously, our concept deserves further investigation.

Excluding TTR, the most commonly measured indicator of the visceral protein compartment is SA, the oldest of all biochemical markers. SA is the

primary determinant of colloid osmotic pressure. It has a low-affinity/high-capacity binding potential for numerous substances, a prolonged biological halflife of ~ 20 days, and a large distribution space (215, 237). Many clinical studies have shown that SA plasma values are correlated to the severity of protein malnutrition and to that of immune deficits, which makes SA an excellent predictive tool of hospital survival (117, 192). However, the clinical usefulness of SA is limited by its inability to identify short-term alterations of nutritional status (75) and by its propensity for extravascular extravasation (82). In contrast, the rapidly turning over TTR, which is less influenced by changes in body fluids than are SA levels (297), allows the daily follow-up of diseased patients. Studies of children with protein-calorie malnutrition in Thailand have shown that TTR falls faster and further from normal than SA during the period of deprivation and is restored to normal more quickly and by lower levels of protein supplementation (1 gm/kg) than SA, which required 4 gm/kg (269). Nonetheless, the idea to combine both SA and TTR markers within a predictive scoring system (134) yielded a faithful bipolar estimate of the visceral protein compartment. This approach is supported by studies showing that the SA and TTR declining or recovering slopes remain positively correlated (86). Their combined measurement therefore provides a more reliable evaluation of the patient's nutritional status (20).

Until now, little attention was paid to the potential usefulness of TTR in assessing the nutritional status of population groups in epidemiological studies. Given the fact that TTR distributes along Gaussian curves (129), a deterioration or improvement of the overall health conditions is accompanied by a shift to the left or to the right of the Gaussian means, with a higher degree of significance than that of most other biochemical and anthropometric measurements of nutritional status (129).

TRANSTHYRETIN IN ACUTE STRESSFUL CONDITIONS

The metabolic consequences of acute stressful conditions have been extensively investigated over the past decades. These disturbances, which include accelerated gluconeogenesis and oxidation of carbohydrates, fatty acid mobilization, and protein synthesis and degradation (97, 168, 315), involve all compartments and functions of the stressed body in several ways. These metabolic changes are distinct from those associated with severe food protein and/or energy restriction alone. Regardless of the causative factor (major injury, trauma, sepsis, burns, cancer cachexia), the stress reaction manifests marked similarities and is typically accompanied by fever, anorexia, muscle wasting, and weight loss. The accompanying hormonal secretory pattern is characterized by rises in the counterregulatory hormones (glucagon, cortisol, adrenaline) and by end-organ insulin resistance (83, 280). Despite quantitative

differences, a qualitative analogy between the reacting responses points to a unifying concept of the stressful condition.

The structural protein pool, mainly in the skeletal musculature, serves as a critical source of AA residues and of metabolic fuel of the stress response. Cuthbertson was the first to show in humans that stress is characterized by a negative N balance (59) whose magnitude is proportionate to the severity of the insult. The classical N balance results from an aggregate of many variables and therefore cannot reveal these specific changes in interorgan substrate fluxes. Nowadays, more sophisticated methods, such as whole-body and specific organ protein turnover methods using labeled tracers, have improved these approaches for dissecting out the various components of whole-body kinetics (324). The current opinion is that, under stressful conditions, protein turnover is stimulated as a result of both augmented tissue proteolysis (mainly in the muscle mass) and specific tissue protein synthesis (mainly in the liver) (143). Protein breakdown releases AA residues, which are preferentially conveyed and incorporated into the hepatic precursor pool involved in the secretion of acute-phase proteins (APPs) and other defense systems (17, 115, 161). However, the rate of protein degradation generally exceeds the rate at which AAs are used for protein synthesis (143), thus yielding a net negative N balance substantiated by rising amounts of urea (59, 179) and of other N catabolites (45) in the urine.

About 15 different mediators, primarily secreted by activated leukocytes and collectively termed cytokines, are implicated in the acute phase response. It was recognized originally that cytokines were responsible for the febrile reaction (69), for muscle proteolysis (50), and for a broad spectrum of synergistic and/or antagonistic effects that may ultimately either reinforce or deteriorate the immune processes of the stressed individual (286). Interleukins 1 and 6 (IL-1, IL-6) and tumor necrosis factor- α (TNF- α) appear to play key roles in the inflammatory reactions. For example, IL-1 (17,500 Daltons as MM) strongly stimulates an increase in whole-body AA flux (319), the oversynthesis of α 1-acid glycoprotein (AGP, orosomucoid) (90), and the early rising production of cortisol by direct adrenotropic (234) or indirect hypophysotropic (21) activation. IL-6 (26,000 Daltons as MM) is the major mediator for the secretion of most other hepatic APPs, especially that of C-reactive protein (CRP) (91). TNF-α (17,000 Daltons as MM) favors muscle proteolysis through the mediation of glucocorticoids (106) as well as glucagon-induced hyperglycemia and AA uptake by the liver (309).

The peak of N urinary excretion culminates within days three to five after the initiation of acute injury (59) and coincides with the nadir recorded for the N negative balance (120) and for the TTR-RBP blood values (151, 225). When the stressful condition subsides—provided that appropriate nutritional support is offered—both N balance (120) and TTR-RBP levels rejoin the physiological

range within a couple of days (133, 225). In contrast, septic and metabolic complications or inadequate dietetic management results in persistent N loss and subnormal TTR-RBP plasma concentrations (120, 133, 321). Thus the evolutionary patterns of N urinary output and of plasma parameters are mirror images of each other, which suggests that the rapidly turning over proteins could eventually reflect the losses of N from the mobilizable endogenous N pools (133). Experimental data using hepatocyte models have confirmed that the molecular alteration triggered by cytokines is situated at transcriptional level, which results in the cytosolic increase of mRNA levels of several APPs, whereas mRNA concentrations of visceral proteins were significantly depressed (66, 187, 190, 252).

The distortion from normal serum values of proteins such as SA and TTR closely parallels the changes in their intrahepatic mRNA levels, which implies that their rates of protein synthesis were reset at new thresholds of priority during the course of inflammation (66, 187, 190, 252). Recent studies have unraveled the molecular mechanisms that control the hepatic production of both inflammatory and visceral proteins (2, 137). The induction of APPs is mediated by an IL-6 nuclear factor (IL-6-NF), which has a high degree of homology to the C/EBF-NF responsible for the expression of visceral markers. Both IL-6- and C/EBP-NFs competitively recognize and promote the same DNA responsive element of the IL-6 gene. Under physiological conditions, in the absence of cytokine stimulation, the visceral indices benefit from preferential production while IL-6-NF is not expressed, which explains the maintenance of APP-mRNAs at low or undetectable cytosolic levels. In contrast, acute stress is characterized by cytokine-induced stimulation of IL-6-NF. which entails a reciprocal shift with enhanced generation of APP-mRNAs and concomitant downregulation of SA- and TTR-mRNAs (2, 137).

At first sight, and from a purely mechanistic point of view, the drop of the short-lived plasma visceral proteins as a result of stress appears to be an inevitable fate poorly modulated by nutritional therapy. Hence, the term "negative APPs" has been suggested in this context to imply that they are inert and passive visceral markers that lose ipso facto any nutritional significance. Ingenbleek has expressed a contrary opinion: that the so-called negative APPs are directly and causally involved in the establishment of all stages of the stress reaction (133). A growing body of recent data suggests that the latter view is especially valid for TTR (133), but we assume that it should be extended to RBP and to CBG. This conclusion is based on the "free hormone hypothesis," which was originally envisaged by Recant & Riggs four decades ago (228), structured within basic considerations by Robbins & Rall (232), and recently extended by Mendel using complex mathematical approaches (178). According to the theory, the following successive major steps should be highlighted: (a) The biological activity of a given hormone or vitamin transported by carrier protein(s) is dependent on its unbound and freely available fraction. (b) Any abrupt decline in the concentration of the carrier protein entails the spontaneous dissociation of the protein-ligand complex following the law of mass action and the dissociation constant (Kd) of the ligand. (c) The delivery of rising amounts of freed ligand molecules into the extracellular spaces proportionally increases their fractional tissue uptake as well as their intracellular disposal rate.

The "free hormone hypothesis" appears to rest on firm ground in the field of both thyroid hormones (175, 176), whose three specific carrier proteins act as buffering reservoirs, thereby allowing the uniform distribution of the hormones among all cells of each irrigated tissue (174). Under steady-state conditions, the minute amounts of free thyroxine (FT4) and free triodothyronine (FT3) available to cells represent only 0.02 and 0.3%, respectively, of the total hormonal concentrations (233). Any acute stressful condition is typically characterized by a "low T3 syndrome" with depressed total and free T3 fractions and elevated reverse T3 (rT3) arising from peripheral impaired monodeiodination of T4 to T3 (131). In contrast, total T4 and TBG values are usually normal or minimally reduced, whereas TTR levels decline as a result of cytokine-induced depressed synthesis (66, 190). This new protein-hormonal equilibrium causes a sudden surge in FT4 concentrations followed by an immediate shift from plasma to tissues (178, 232).

The liver, which may harbor in a nonphysiologically active form as much as 40% of the total extrathyroidal T4 reserves (44), may contribute to the feeding of the FT4 pool (112) instantly taken up by cells before feedback mechanisms can intervene. Under these circumstances, most clinical studies have documented that TSH remains unaffected, with FT4 levels situated at the upper range of normalcy or slightly elevated (221, 284). However, direct plasma measurement largely underscores the magnitude of cellular impregnation by FT4. The increased urinary excretion of FT4 (225) certainly provides a better estimate of its increased extracellular availability. From the thyroid function point of view, acute stress is thus a very complex and dichotomic situation characterized by an overall downregulation of organs not directly involved in the stress reaction. This observation correlates with the relative refractoriness of T3 cytosolic and nuclear binding sites of these organs, which teleologically seek to spare the N body reserves (300). In contrast, the liver and other tissues, such as white blood cells (317) (which actively contribute to defense systems, immune responses, and wound healing), reveal strong overstimulation. The peripheral turnover of thyroid hormones may be augmented severalfold in bacterial sepsis (104) and parasitic infestation (310), an outcome consistent with their increased consumption and thermogenic effects (17, 168) at cellular level. The balance between these two down- and upregulated influences presumably depends on the nature of pathogens and the proportion of affected cells in the stressed body, which explains the disparity of metabolic responses.

Recent years have witnessed substantial advances in knowledge of the multifaceted actions of the thyroid gland. Most authors agree with the view that the FT4 concentrations in the extracellular fluids appear to be the principal determinant of peripheral thyroid status, thus ensuring the fine-tuning retrocontrol of the endocrine organ secretory rate (131). In contrast, intracellular FT3, whose bulk is generated by tissue FT4 monodeiodination, appears to be the major metabolically active hormonal compound (131, 210). The cellular sites of actions for thyroid stimuli remain a field of intense research. Although not excluded, the previously proposed cytosolic targets, such as plasma membrane, enzymes, or mitochondrion, have lost some consistency in recent years (210). Some cytosolic sites of action have been confirmed (166), but most surveys contend that the main hormonal effects are situated at nuclear level and related to the selective regulation of specific genes (210). The occupancy of hepatic nuclear receptors by FT3 is a direct and very rapid second-order process (209). The radioautographic study of the two-dimensional activity profile of mRNA translational products indicates that some of them are stimulated while others are depressed, according to the thyroid hormonal status (251). This dualistic expression of hepatic genes was confirmed recently in a human hepatoma cell-line culture, thus revealing an increase in AGP-mRNA levels that contrasts with the decrease in TBG-mRNA values (158). In such hepatocyte models, the saturation kinetics of T3 nuclear receptors may be reached in less than 2 min (209), causing a significant elevation of the AGPmRNA concentrations as early as 30 min after the addition of T3 to the incubation medium (156).

Under steady-state conditions, the plasma free retinol fraction does not exceed 5% of its totally bound concentration (100). During the onset of acute stress, the drop of RBP parallels that of its counterpart protein (151, 225). As shown for TTR and T4, the new RBP-retinol mass equilibrium probably causes release into the extracellular space of increasing amounts of free vitamin becoming available to cells. Once again, the kidneys prevent excessive accumulation of the potentially toxic free retinol (225), whose urinary leakage reflects the augmented extracellular pool.

Vitamin A is required for vision, growth, and reproduction as well as for the maintenance of differentiated epithelia and mucus secretion (27, 99, 217, 227). Moreover, vitamin A is deeply involved in the activation and modulation of the host defense systems (64, 318). However, interpretation of these aspects is complex and must be cautious because *all-trans*-retinol undergoes oxidation to form *all-trans* retinoic acid, the principal ligand to the vitamin A-related genomic receptors. In addition, other retinoids sharing with retinol common functional properties except in vision and spermatogenesis are formed. The

delivery to target organelles of these retinoid compounds endowed with high morphogenic and cytotoxic potency is finely regulated by several membrane, cytosolic, and nuclear-specific receptors (167, 206). Nevertheless, rapid and spontaneous dissociation of retinol from plasma RBP has been reported (204). Using cultured animal keratinocytes, the uptake of freed retinol is many times more rapid than the release of retinol by cell-surface receptors for RBP (57). Moreover, maximal intracellular accumulation and biological effects of free retinol delivered to human keratinocytes are recorded after 3 h, in contrast to the very slow results obtained with holo-RBP (58). The free hormone/vitamin hypothesis predicts that retinol released in free form would favor rapid transmembrane entry under acute stressful conditions.

Vitamin A compounds are thought to modulate the activity of several factors that mediate growth and immunity by two distinct major pathways (62). The first is cytosolic and is situated at ribosomal level, where retinoids promote the incorporation of carbohydrate residues into native glycopeptidic chains. The second is nuclear and relates to the description of at least three specific binding sites (159) controlling the transcription of genomic products. For example, physiological concentrations of vitamin A stimulate the production of interleukins (288), proteins (110), APPs (13), and enzymatic molecules involved in the host defense systems (49, 135). Conversely, vitamin A deficiency is responsible for the defective glycosylation of two rat APPs, α 1-macroglobulin (154) and fibronectin (155). Like thyroid hormones, vitamin A supplementation may exhibit ambivalent properties and depress the synthesis of secretory products such as interferon- γ (43).

The collective data strongly suggest that both TTR and RBP, as a result of acute suppression of their hepatic synthesis, actively participate in the onset of stress reactions and therefore deserve to be described as "acute-booster reactants" (ABRs). The delivery of free thyroid and retinoid compounds, readily available to cells by simple diffusion, makes it possible to initiate the positive and negative regulatory controls associated with injury. The magnitude and duration of this transient hyperthyroid and hyperretinoid status depend on the decrement between pre- and poststress plasma TTR-RBP levels undergoing spontaneous dissociation and freeing all of its bound ligands. This release persists as long as TTR-RBP drops and appears to be proportionate to the severity of stress and to the degree of negative N balance. However, initiation of these inducing steps does not exceed the four or five days necessary to reach the nadir of TTR-RBP, which coincides with the peak of urinary excretion of N catabolites.

These results explain why an optimal nutritional status, as assessed by TTR-RBP levels within the normal range, confers to the stressed body the metabolic advantage of developing appropriate responses of higher magnitude and longer duration. Conversely, a preceding nutritional status characterized

by deterioration and subnormal TTR-RBP plasma levels might be accountable for an overall impairment of the aforementioned thyroid- and retinoid-dependent processes in infected or stressed PEM subjects. These abnormalities are likely to aggravate and to interact with the other multiple immune deficiencies directly generated by the inhibition of cytokines in protein malnutrition (22, 157). The data argue strongly for early recognition and correction of PEM states by appropriate nutritional management in order to help the stressed body surmount the harmful consequences of malnutrition. Clinicians must be aware that anthropometry greatly underestimates the importance of protein depletion and that biochemical parameters constitute earlier and more useful indices for valid nutritional assessment and therapeutic purposes.

TRANSTHYRETIN IN CHRONIC STRESSFUL DISORDERS

The complex variety of metabolic effects induced by thyroid hormones and retinol during acute stages of stress cannot be dissociated from those triggered by cortisol. These active compounds are transported by specific carrier proteins and exhibit similarities in their evolutionary patterns during the stress response. Once released in free form into the bloodstream, physiologically active T4, retinol, and cortisol may be taken up by peripheral cells, a process that entails a cascade of intracellular reactions ultimately regulated by the same superfamily of nuclear receptors (205). Nevertheless, cortisol has additional specific activities aimed at strengthening the stress reaction and preventing undesirable consequences, which explains why it is predominantly operative in chronic situations.

CBG is a glycoprotein (42,650 Daltons as MM) secreted by the liver (312). It has a unique binding site for cortisol and conveys ~ 90% of the circulating cortisol. Under normal circumstances, this protein-hormonal complex stands in equilibrium with ~ 4% of physiologically active free cortisol (261, 282). In contrast to TTR and RBP, CBG plasma concentrations do not show differences between the sexes and are only modestly influenced by dietary manipulation (8). However, like TTR and RBP, CBG suffers an acute suppression in its hepatic synthesis during inflammatory disorders (223, 327). The sharp decline in CBG plasma levels thus coincides with the aforementioned cytokine-induced overstimulation of both pituitary and adrenal glands (21, 234), which leads to increased rates of cortisol production (14). Owing to the strikingly divergent alterations observed for each component of the carrier-ligand system, a greatly increased proportion of free cortisol is released into the extracellular space, where it is readily available to cells. The enhanced supply of unbound hormone appears to be correlated to the severity and duration of the stressful condition (17, 280) and reaches many times the normal tissue requirements (316). The direct measurement of cortisoluria also reveals greatly augmented values (14), which reflect, once again, the enlarged circulating free pool and kidney overflow (184). This intense hormonal response does not exceed a few days and subsides after one week in elective surgery (184) but may be sustained for a prolonged period in situations characterized by chronicity, complications, and/or relapses, e.g. burns (316). The latter clinical context is typically delineated by a further glucocorticoid-induced depression in CBG plasma values (248), which implies that chronic inflammatory disorders are evolving towards a generalized, self-sustained stage of hypercorticism.

Because rapid plasma dissociation and liver unidirectional influx rates have been reported for cortisol (282), it seems likely that the free hormone hypothesis could apply to this compound (177). The metabolic effects of cortisol are mediated by the activation of adenylate cyclase (235) and/or by the binding to cytosolic and nuclear receptors (238). The dual mode of action already reported for thyroid (158, 251) and retinoid (43, 288) molecules also characterizes glucocorticoid activity, which may boost the production of some APPs (such as AGP) (298) or enzymes (such as tyrosine aminotransferase and alkaline phosphodiesterase I) (238). Conversely, glucocorticoids may exhibit wellestablished negative regulatory capacities (1, 279). However, there is a striking difference between thyroid/retinoid components, which operate as true inducers or repressors of the stress reaction, and steroid hormones, which behave as secondary modulators of cellular activity. This difference stems from experimental studies in which the enhanced de novo accumulation of AGPmRNA and of some other APPs also develops in adrenalectomized rats after turpentine-induced inflammation (16). This investigation demonstrates that steroid-dependent processes require primary effector systems—likely thyroid hormones in the case of AGP response (158). This cascade of molecular events is consistent with the general concept that glucocorticoids function as modulating agents of primary transcripts (238).

In healthy individuals undergoing stress, steroidogenesis is fairly well adapted to meet the increased metabolic demands of the stressed body (301). This response seems to fulfill teleological purposes and is vital for survival because both excessive and defective pituitary-adrenocortical responses have adverse prognosis significance (259). This view is corroborated by the discovery of a structural analogy between CBG and several components of the serine-proteinase inhibitor (serpin) superfamily, notably with TBG and α 1-protease inhibitor (A₁-PI) (107). Both CBG and A₁-PI interact specifically and offer excellent substrates for neutrophil elastase on the surface of activated macrophages (109). Enzymatic cleavage of CBG leads to conformational changes and disruption of its steroid binding site, followed by a 10-fold drop in the Ka of cortisol (213). This process should favor the selective delivery of cortisol at the site of inflammation (108), thus locally

reinforcing the hypercortisolemic background of the injured tissues. Glucocorticoids play simultaneously different roles in preventing healthy tissues from the destructive potential of cytokines (70, 289) and from the overactivity of nitric oxide synthase (92) exacerbated in inflammatory disorders (181). Depending on their concentration, duration of cell exposure, and metabolic environment, the net biological effects of cytokines may ultimately exert beneficial or deleterious effects on the host (286). The inactivation of A₁-PI and of other serpin components, which represent more than 10% of the total protein in human plasma (287), allow the serine proteinases to operate more efficiently in the inflammatory context and thereby contribute to the preservation of N endogenous pools triggered by T3 downregulation (300). The low TBG plasma values characterizing seriously ill patients (114) might well result from the combined effects of both defective synthesis (126, 158) and enzymatic cleavage (213) and thus contribute to increased turnover and disposal of thyroid hormones in PEM (132) and inflammatory states (104, 310).

In these complex chronic situations in which the detrimental effects of both malnutrition and stress are joined, the drop in visceral proteins may be regarded as the sum of both nutritional and inflammatory injuries. In this context, no single parameter fulfills the scoring task, and the correct appraisal of each causal factor requires a simultaneous measurement of indices reflecting all of them (134). In an attempt to encompass these two interwoven aspects of the disease spectrum, a predictive scoring system has been proposed that would allow (regardless of sex and age) the stratification of critically ill patients by risk of complications or death (134).

$$PINI = \frac{CRP (mg/liter) \times AGP (mg/liter)}{SA (g/liter) \times TTR (mg/liter)}$$

Thus this prognostic inflammatory and nutritional index (PINI) is a quotient that aggregates the product of the most reliable indicators of protein status with that of CRP and AGP. These last two APPs were selected after stepwise discriminant analysis of seven inflammatory markers clinically employed in current use (134). The PINI scoring system yields values below 1.0 under normal circumstances but that rise above this baseline as the overall health status deteriorates. Values exceeding a threshold of ~30 reflect severely morbid and/or life-threatening conditions. The choice of both CRP and AGP was largely due to their biological complementarity, which ensures the broadest coverage of the inflammatory reaction. On the one hand, AGP is one of the most glycosylated of all known glycoproteins, and its hepatic overproduction is predominantly set in motion by IL-1 (90). AGP may be regarded as a slow-reacting marker ($T_{1/2} = 5-7$ days) whose intracellular maturation process necessitates a reduction in size of the precursor form (257), which yields a

primary transcript characterized by increased stability (298). These events may be influenced by thyroid (158) and steroid (298) hormones. On the other hand, CRP is an unglycosylated pentaxin barely detectable in normal blood and whose hepatic synthesis is mainly initiated by IL-6 (91). This APP is an exceedingly fast responder ($T_{1/2} = 4-6$ h), and its production reportedly is facilitated in the course of the stress reaction (169). On the basis of available molecular data, we assume that this enhanced secretory process of CRP should be inversely correlated to the drop in ABR levels and therefore proportionate to the amount of freed ligand(s). It seems likely that these released mediators would act as repressors of the inhibitory control of CRP secretion (but not of CRP synthesis) (169).

Within the PINI formula, CRP-TTR and AGP-SA appear as rapid and slow reacting couples, respectively. They respond with specific kinetics along differentiated pathways to IL-1 and IL-6, which minimizes the risk of yielding a prognostic value that does not concord with the severity of the clinical condition. These response patterns are all the more important because, in contrast to the mirror image of changes in CRP-AGP vs SA-TTR levels during acute stress, each component of the PINI may evolve in a unique and independent way under chronic circumstances, in agreement with the view that immunological and nutritional indices move along uncorrelated pathways (101). This process is particularly well-documented in cancer patients in whom N balance and visceral indicators may be significantly improved as a result of appropriate refeeding, notwithstanding unchanged neoplastic burden (230). In contrast, the correct follow-up of inflammatory parameters not accompanied by that of visceral proteins incurs the risk of inadequate nutritional support and of undesirable complications. That the magnitude of the acute phase protein response may be depressed by protein deficiency both in experimental (144) and clinical (71) surveys is well established. The description of significantly lower CRP and AGP in surgical patients, in whom deep sternal infection manifests later (180), is strongly indicative of unrecognized preexisting malnutrition.

The PINI formula is presently employed on a large scale and under validation in various diseased states, notably those affecting children (222), elderly persons (4, 219), and burned (160) and trauma (299) patients. The predictive scoring system is not intended to establish any precise diagnosis; rather, its nonspecificity permits its broad applicability in most clinically significant stressful states. Advantages and limitations of this approach remain to be determined. However, the available data clearly show that the PINI system fulfills its most promising potential in identifying subclinical or marginal situations as well as complex inflammatory and nutritional disorders in protracted illnesses. It thus represents a major improvement over normal routine laboratory methods.

SUMMARY

The name "transthyretin" reflects the dual physiological roles of this tetrameric unglycosylated plasma protein. TTR is one of three specific carrier proteins involved in the transport of both thyroid hormones and of retinol through the mediation of RBP. TTR is a product of the visceral compartment, and its hepatic synthesis is exquisitely sensitive to both the adequacy and levels of protein and energy intakes—hence the proposal of TTR as a nutritional marker. To date, 38 TTR variants have been described, most of which are associated with variable degrees of cardiac and/or neural tissue amyloid deposits. All known variants arise from a single AA substitution due to single point mutation in the coding region of the TTR gene. Under acute stress conditions, the synthesis of TTR, RBP, and CBG is abruptly depressed by a cytokine-directed orchestration of new metabolic priorities, with a redistribution of organ and tissue protein pools. It is proposed that TTR, RBP, and CBG behave as acute-booster reactants (ABRs), actively participating in the cascade of metabolic events characterizing the stress reaction along pathways best explained by the free hormone/vitamin hypothesis. The latter is governed by the law of mass action—the spontaneous dissociation and instant uptake by hepatocytes of the ligands freed from their specific carrier proteins, which creates a transient hyperthyroid, hyperretinoid, and hypercortisolic climate. This response generally does not exceed four or five days because the initial impact of injury normally subsides, but it may last longer if complications occur. The magnitude and adequacy of the stress responses depend on the preceding nutritional status as assessed by TTR plasma levels and are proportionate to the severity of insult. Clinical, animal, and molecular studies concur to demonstrate the dualistic stimulatory or inhibitory effects triggered by the ligands, whose unmetabolized fractions are excreted in the urinary output. Thyroid hormones and retinoids appear to control the early maturation processes and the synthesis of primary transcripts, whereas cortisol preferentially modulates the secondary responses and confers a protective effect on healthy tissues. During acute stress, the evolutionary patterns of visceral proteins and inflammatory markers exhibit compulsory mirror images. However, they change in independent ways under more chronic circumstances. A relatively simple biochemical micromethod based on the simultaneous measurement of plasma TTR, albumin, CRP, and orosomucoid aggregated into a PINI is proposed for the early recognition and follow-up of both nutritional and inflammatory facets of the disease spectrum.

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Literature Cited

- Akerblom IE, Slater EP, Beato M, Baxter JD, Mellon P. 1988. Negative regulation by glucocorticoids through interference with a cAMP responsive enhancer. Science 241:350-53
- Akira S, Isshiki H, Nakajima T, Kinoshita S, Nishio Y, et al. 1992. Regulation of expression of the interleukin 6 gene: structure and function of the transcription factor NF-IL6. In Polyfunctional Cytokines: Il-6 and LiF, 167:47-67. Ciba Found. Symp. Chichester, U.K.: Wiley
- Aleshire SL, Bradley C, Richardson L, Parl F. 1983. Localisation of human prealbumin in choroid plexus epithelium. J. Histochem. Cytochem. 31:608– 12
- Alix E, Papin A, Fromont P, Queinec S, Vieron MC, et al. 1993. Index pronostique nutritionnel et inflammatoire (PINI): évaluation en court séjour gériatrique chez 260 personnes âgées de plus de 75 ans. Age Nutr. 4:63-71
- Almeida MR, Altland K, Rauh S, Gawinowicz MA, Moreira P, et al. 1991. Characterization of a basic transthyretin variant -TTR Arg 102- in the German population. Biochim. Biophys. Acta 1097:224-26
- Almeida MR, Hesse A, Steinmetz A, Maisch B, Altland K, et al. 1991. Transthyretin Leu 68 in a form of cardiac amyloidosis. Basic Res. Cardiol. 86: 567-71
- Almeida MR, Ferlini A, Forabosco A, Gawinowicz MA, Costa PP, et al. 1992. Two transthyretin variants (TTR Ala-49 and TTR Gln-89) in two Sicilian kindreds with hereditary amyloidosis. Hum. Mutat. 1:211-15
- Anderson KE, Rosner W, Khan MS, New M, Pang S, et al. 1987. Diet-hormone interactions: Protein/carbohydrate ratio alters reciprocally the plasma levels of testosterone and cortisol and their respective binding globulins in man. Life Sci. 40:1761-68
- Andrade C. 1952. A peculiar form of peripheral neuropathy. Familial atypical generalized amyloidosis with special involvement of peripheral nerves. *Brain* 75:408-22
- Andrea TA, Cavalieri RR, Goldfine ID, Jorgensen EC. 1980. Binding of thyroid hormones and analogues to the human plasma protein prealbumin. *Biochemis*try 19:55-63
- Andreoli M, Robbins J. 1962. Serum proteins and thyroxine protein interac-

- tion in early human fetuses. J. Clin. Invest. 41:1070-77
- Bach JF. 1983. Thymulin (FTS-Zn). Clin. Immunol. Allergy 3:133-56
- Barber EF, Cousins RJ. 1987. Induction of ceruloplasmin synthesis by retinoid acid in rats: influence of dietary copper and vitamin A status. J. Nutr. 117:1615– 22
- Barton RN, Weijers JW, Horan MA. 1993. Increased rates of cortisol excretion in elderly women 2 weeks after proximal femur fracture. Eur. J. Clin. Invest. 23:171-76
- Bates J, McClain CJ. 1981. The effect of severe zinc deficiency on serum levels of albumin, transferrin and prealbumin in man. Am. J. Clin. Nutr. 34:1655-60
- Baumann H, Firestone GL, Burgess TL, Gross K, Yamamoto K, Held WA. 1983. Dexamethasone regulation of α₁-acid glycoprotein and other acute phase reactants in rat liver and hepatoma cells. J. Biol. Chem. 258:563-70
- Beisel W. 1975. Metabolic response to infection. Annu. Rev. Med. 26:9-20
- infection. Annu. Rev. Med. 26:9-20
 18. Benson MD II, Turpin JC, Lucotte G, Zeldenrust S, Le Chevallier B, Benson MD. 1993. A transthyretin variant (alanine 71) associated with familial polyneuropathy in a French family. J. Med. Genet. 30:120-22
- Bernard C. 1865. Introduction à l'Etude de la Médecine Expérimentale, pp. 1– 400. Paris: Baillière
- Bernstein LH, Leukhardt-Fairfield CJ, Pleban W, Rudolph R. 1989. Usefulness of data on albumin and prealbumin concentrations in determining effectiveness of nutritional support. Clin. Chem. 35: 271-74
- Bernton EW, Beach JE, Holaday JW, Smallridge R, Fein HG. 1987. Release of multiple hormones by a direct action of interleukin-1 on pituitary cells. Science 238:519-21
- Bhaskaram P, Sivakumar B. 1986. Interleukin-1 in malnutrition. Arch. Dis. Child. 61:182-85
- Bhatia J, Ziegler EE. 1983. Retinolbinding protein and prealbumin in cord blood of term and preterm infants. Early Hum. Dev. 8:129-33
- Bistrian BR, Sherman M, Young VR. 1981. The mechanisms of nitrogen sparing in fasting supplemented by protein and carbohydrate. J. Clin. Endocrinol. Metab. 53:874-78
- Blake CCF, Geisow M, Oatley S, Rerat B, Rerat C. 1978. Structure of prealbu-

- min: secondary, tertiary and quaternary interactions determined by Fourier refinement at 1.8 Å. J. Mol. Biol. 121:339–56
- Blake CCF, Oatley S. 1977. Protein-DNA and protein-hormone interactions in prealbumin: a model of the thyroid hormone nuclear receptor? Nature 268: 115-20
- Blaner WS. 1989. Retinol-binding protein: the serum transport protein for vitamin A. Endocr. Rev 10:308-16
- Bleiberg F, Vranckx R, Wade S, Nunez EA. 1985. A simplified method for the purification of rat thyroxine-binding prealbumin. Factors influencing its circulating level. Biochim. Biophys. Acta 828:270-77
- Bleiberg-Daniel F, Le Moullac B, Maire JC, Wade S. 1990. Failure of tryptophan deficiency to reduce specifically serum levels of transthyretin or albumin in rats. J. Nutr. 120:1610-16
- Branch WT, Robbins J, Edelhoch H. 1971. Thyroxine-binding prealbumin. Conformations in aqueous solutions. J. Biol. Chem. 246:6011-18
- Branch WT, Robbins J, Edelhoch H. 1972. Thyroxine-binding prealbumin. Conformation in urea and guanidine. Arch. Biochem. Biophys. 152:144-51
- Braverman LE, Dawber NA, Ingbar SH. 1966. Observations concerning the binding of the thyroid hormones in sera of normal subjects of varying age. J. Clin. Invest. 45:1273-79
- Braverman LE, Ingbar SH. 1967. Effects of norethandrolone on the transport in serum and peripheral turnover of thyroxine. J. Clin. Endocrinol. Metab. 27: 389-96
- Brennan MF, Fitzpatrick GF, Cohen KH, Moore FD. 1975. Glycerol: major contributor to the short term protein sparing effect of fat emulsions in normal man. Ann. Surg. 182:386-94
- Burri BJ, Niedlinger TR, Vanloan M, Keim NL. 1990. Effect of low-calorie diets on plasma retinol-binding protein concentrations in overweight women. J. Nutr. Biochem. 1:484-86
- Burri BJ, Kutnink MA, Neidlinger TR. 1991. Assay of human transthyretinbound holo-retinol-binding protein with reversed-phase high-performance liquid chromatography. J. Chromatogr. 567: 369-80
- Burton PM, Iden S, Mitchell K, White A. 1978. Thymic hormone-like restoration by human prealbumin of azathioprine sensitivity of spleen cells from thymectomized mice. Proc. Natl. Acad. Sci. USA 75:823-27

- Burton PM, Horner BL, Jones G, Lin T, Nestor J, et al. 1987. Immunoenhancing activity of the amino-terminal domain of human prealbumin: isolation, characterization and synthesis. Int. J. Immunopharmacol. 9:297-305
- Cahill GF Jr, Herrera MG, Morgan AP, Soeldner JS, Steinke J, et al. 1966. Hormone-fuel interrelationships during fasting. J. Clin. Invest. 45:1751-69
- Calloway DH, Spector H. 1954. Nitrogen balance as related to caloric and protein intake in active young man. Am. J. Clin. Nutr. 2:405-12
- Cano N, di Costanzo-Dufetel J. 1987.
 Transthyrétine (Préalbumine) sérique.
 Nutr. Clin. Métab. 1:7-15
- Cano N, di Costanzo-Dufetel J, Calaf R, Durbec JP, Lacombe P, et al. 1988. Prealbumin-retinol binding protein-retinol complex in hemodialysis patients. Am. J. Clin. Nutr. 47:664-67
- Carman JA, Hayes CE. 1991. Abnormal regulation of IFN-γ secretion in vitamin A deficiency. J. Immunol. 147:1247-52
- Cavalieri RR, Searle GL. 1966. Role of distribution between plasma and liver of ¹³¹I-labeled L-thyroxine in man: observations of subjects with normal and decreased serum thyroxine-binding globulin. J. Clin. Invest. 45:939-49
- Cederblad G, Schildt B, Larsson J, Liljedahl SO. 1983. Urinary excretion of carnitine in multiply in jured patients on different regimens of total parenteral nutrition. *Metabolism* 32:383-89
- Chandra RK. 1979. Serum thymic hormone activity in protein-energy malnutrition. Clin. Exp. Immunol. 38:228-30
- Cheng SY, Pages RA, Saroff HA, Edelhoch H, Robbins J. 1977. Analysis of thyroid hormone binding to human serum prealbumin by 8-anilino naphtalene-1-sulfonate fluorescence. Biochemistry 16:3707-13
- Cheung CK, Swaminathan R. 1989. Automated immunoturbidimetric methods for the determination of retinol-binding protein, prealbumin and transferrin in urine. Clin. Biochem. 22:425-27
- Chiocca EA, Davies PJ, Stein J. 1988. The molecular basis of retinoic acid action. Transcriptional regulation of tissue transglutaminase gene expression in macrophages. J. Biol. Chem. 263: 11584-89
- Clowes GHA Jr, George BC, Villee CA Jr, Saravis CA. 1983. Muscle proteolysis induced by a circulating peptide in patients with sepsis and trauma. New Engl. J. Med. 308:545-52
- Collins VP, Jacobsson B, Pettersson T, Carlstrom A. 1986. Monoclonal anti-

- bodies to transthyretin, Scand. J. Clin. Lab. Invest. 46:761-69
- Cornwell GG III, Murdoch W, Kyle RA, Westermark P, Pitkanen P. 1983. Frequency and distribution of senile cardiovascular amyloid. A clinicopathologic correlation. Am. J. Med. 75:618

 23
- Costa PP, Figueira A, Bravo F. 1978. Amyloid fibril protein related to prealbumin in familial amyloidotic polyneuropathy. Proc. Natl. Acad. Sci. USA 75:4449-503
- Costa RH, Lai E, Darnell JE. 1986. Transcriptional control of the mouse prealbumin (transthyretin) gene: Both promotor sequences and a distinct enhancer are cell specific. Mol. Cell. Biol. 6:4697– 708
- Costa RH, Lai E, Grayson DR, Darnell JE. 1987. The cell-specific enhancer of the mouse transthyretin (prealbumin) gene binds a common factor at one site and a liver-specific factor(s) at two other sites. Mol. Cell. Biol. 8:81-90
- Cowley DM, Davey J, Hjelm NM. 1983.
 A radioimmunoassay of human prealbumin in body fluids. Clin. Chim. Acta 134:69-76
- Creek KE, Silverman-Jones CS, De Luca LM. 1989. Comparison of the uptake and metabolism of retinol delivered to primary mouse keratinocytes either free or bound to rat serum retinol-binding protein. J. Invest. Dermatol. 93:283– 89
- Creek KE, St. Hilaire P, Hodam JR. 1993. A comparison of the uptake, metabolism and biologic effects of retinol delivered to human keratinocytes either free or bound to serum retinol-binding protein. J. Nutr. 123(Suppl. 2):356-61
- Cuthbertson DP. 1930. The disturbance of metabolism produced by bony and nonbony injury with notes on certain abnormal conditions of bone. *Biochem.* J. 24:1244-63
- Davis PJ, Handwerger BS, Gregerman R. 1972. Thyroid hormone binding by human serum prealbumin (TBPA). Electrophoretic studies of triiodothyronine-TBPA interaction. J. Clin. Invest. 51: 515-21
- de Jong FA, Schreiber G. 1987. Messenger RNA levels of plasma proteins in rat liver during protein depletion and refeeding. J. Nutr. 117:1795-800
- De Luca LM. 1991. Retinoids and their receptors in differentiation, embryogenesis and neoplasia. FASEB J. 5: 2924– 33
- De Nayer P, Van den Schrieck H, Koch M, De Visscher M. 1966. A simplified

- method for the preparation of thyroxine-binding prealbumin. *Biochim. Bio*phys. Acta 124:411-12
- Dennert G. 1984. Retinoids and the immune system: immunostimulation by vitamin A. In *The Retinoids*, ed. MB Sporn, AB Robert, DS Goodman, 2: 373-90. New York: Academic
- de Vera N, Cristofol R, Rodriges Farre E. 1988. Protein binding and stability of norepinephrine in human blood plasma. Involvement of prealbumin, alpha 1-acid glycoprotein and albumin. Life Sci. 43:1277-86
- Dickson PW, Howlett GJ, Schreiber G. 1982. Metabolism of prealbumin in rats and changes induced by acute inflammation. Eur. J. Biochem. 129:289-93
- Dickson PW, Howlett G, Schreiber G. 1985. Rat transthyretin (prealbumin). Molecular cloning, nucleotide sequence, and gene expression in liver and brain. J. Biol. Chem. 260:8214-19
- 68. Dickson PW, Aldred AR, Marley P, Bannister D, Schreiber G. 1986. Rat choroid plexus specializes in the synthesis and the secretion of transthyretin (prealbumin). Regulation of transthyretin synthesis in choroid plexus is independent from that of liver. J. Biol. Chem. 261:3475-78
- Dinarello CA. 1979. Production of endogenous pyrogen. Fed. Proc. 38:52-56
- Doherty GM, Jensen J, Buresh C, Norton JA. 1992. Hormonal regulation of inflammatory cell cytokine transcript and bioactivity production in response to endotoxin. Cytokine 4:55-62
- Doherty JF, Golden MHN, Raynes JG, Griffin GE, McAdam KP. 1993. Acutephase protein response is impaired in severely malnourished children. Clin. Sci. 84:169-75
- Dwulet FE, Benson MD. 1983. Polymorphism of human thyroxine prealbumin. Biochim. Biochys. Res. Commun. 114:657-62
- Dwulet FE, Benson MD. 1986. Characterization of a transthyretin (prealbumin) variant associated with familial amyloidotic polyneuropathy type II (Indiana/Swiss). J. Clin. Invest. 78:880-86
- Episkopou V, Maeda S, Nishigushi S, Shimada K, Gaitanaris GA, et al. 1993. Disruption of the transthyretin gene results in mice with depressed levels of plasma retinol and thyroid hormone. Proc. Natl. Acad. Sci. USA 90:2375-79
- Erstad BL. 1992. Serum albumin concentrations: Who needs them? Ann. Pharmacother. 26:1134-38
- 76. FAO/WHO/UNU. 1985. Energy and Protein Requirements. Rep. of a Joint

- FAO/WHO/UNU Consultation. Techn. Rep. Ser. 724. Geneva: WHO
- Felding P, Fex G. 1983. Factors responsible for the decreased plasma concentration of prealbumin during acute inflammation and fasting in the rat. Acta Physiol. Scand. 117:377-83
- Ferguson RN, Edelhoch H, Saroff HA, Robbins J. 1975. Negative cooperativity in the binding of thyroxine to human serum prealbumin. Preparation of tritium-labeled 8-anilino-1-naphtalene sulfonic acid. Biochemistry 14:282-89
- Ferlini A, Salvi F, Patrosso C, Fini S, Vezzoni P, Forabosco A. 1993. Gly 47 Ala: A new transthyretin gene mutation in hereditary amyloidosis TTR-related. J. Rheumatol. 20:187
- Fitch NJ, Akbari MT, Ramsden DB. 1991. An inherited non-amyloidogenic transthyretin variant, [Ser⁶]-TTR, with increased thyroxine-binding affinity, characterized by DNA sequencing. J. Endocrinol. 129:309-13
- Fjeld CR. 1991. Control of protein synthesis and its relationship to the bioenergetics of growth. Acta Paediatr. Scand. S374:15-21
- Fleck A, Raines G, Hawker F, Trotter J, Wallace PI, et al. 1985. Increased vascular permeability: a major cause of hypoalbuminemia in disease and injury. Lancet 1:781-84
- Frayn KN. 1986. Hormonal control of metabolism in trauma and sepsis. Clin. Endocrinol. 24:577-99
- Freeman JB, Stegink LD, Wittine M, Danney MM, Thompson RG. 1977. Lack of correlation between nitrogen balance and serum insulin levels during protein sparing with and without dextrose. Gastroenterology 73:31-36
- Fries E, Gustafsson L, Peterson PA. 1984. Four secretory proteins synthesized by hepatocytes are transported from endoplasmic reticulum to Golgi complex at different rates. EMBO J. 3:147-52
- Fulop T, Herrmann F, Rapin CH. 1991. Prognostic role of serum albumin and prealbumin levels in elderly patients at admission to a geriatric hospital. Arch. Gerontol. Geriatr. 12:31-40
- Fürst P. 1985. Regulation of intracellular metabolism of amino acids. In Nutrition in Cancer and Trauma Sepsis, ed. D Bozzetti, pp. 21-35. Karger: Basel
- Gavrilesco K, Courcon J, Hillion P, Uriel J, Lewin J, Grabar P. 1955. A study of normal human cerebrospinal fluid by the immunoelectrophoretic method. Nature 176:976
- 89. Gebre-Medhin M, Ewal U, Fuvemo T.

- 1985. Reduced serum proteins in diabetic children on a twice-daily insulin schedule. Acta Paediatr. Scand. 74:961-65
- Geiger TH, Andus T, Klapproth J, Northoff H, Heinrich PC. 1988. Induction of alpha 1-acid glycoprotein by recombinant human interleukin-1 in rat hepatoma cells. J. Biol. Chem. 263: 7141-46
- Geiger TH, Andus T, Klapproth J, Hirano T, Kishimoto T, Heinrich PC. 1988. Induction of rat acute-phase proteins by interleukin 6 in vivo. Eur. J. Immunol. 18:717-21
- Geller DA, Nussler AK, di Silvio M, Lowenstein CJ, Shapiro RA, et al. 1993. Cytokines, endotoxin, and glucocorticoids regulate the expression of inducible nitric oxide synthase in hepatocytes. Proc. Natl. Acad. Sci. USA 90: 522-26
- Georgieff MK, Sasanow SR, Mammel M, Ophoven J, Pereira GR. 1986. Cord prealbumin values in newborn infants: effect of prenatal steroids, pulmonary maturity, and size for dates. J. Pediatr. 108:972-76
- Gitlin D, Kumate J, Urrusti J, Morales C. 1964. The selectivity of the human placenta in the transfer of plasma proteins from the mother to fetus. J. Clin. Invest. 43:1938-51
- Glover J, Jay C, White GH. 1974. Distribution of retinol-binding protein in tissues. Vitam. Horm. 32:215-35
- Goldsmith BM, Munson S. 1987. Rate nephelometry and radial immunodiffusion compared for measuring serum prealbumin. Clin. Chem. 33:161-63
- Goldstein SA, Elwyn DH. 1989. The effects of injury and sepsis on fuel utilization. Annu. Rev. Nutr. 9:445-73
- 98. Goodman DS, Peters T, Robbins J, Schwick G. 1981. Prealbumin becomes transthyretin. Nomenclature Committee—IUB and JCBN Newsletter. J. Biol. Chem. 256:12-14
- Goodman DS. 1984. Plasma retinolbinding protein. In *The Retinoids*, ed. MB Sporn, AB Roberts, DS Goodman, 2:41-52. New York: Academic
- Goodman DS. 1984. Vitamin A and retinoids in health and disease. New Engl. J. Med. 310:1023-31
- Goodwin JS, Garry PJ. 1988. Lack of correlation between indices of nutritional status and immunologic function in elderly humans. J. Gerontol. 43:M46– 49
- Gorevic P, Prelli F, Wright J, Pras M, Frangione B. 1989. Systemic senile amyloidosis. Identification of a new pre-

- albumin (transthyretin) variant in cardiac tissue: immunologic and biochemical similarity to one form of familial amyloidotic polyneuropathy. J. Clin. Invest. 83:836-43
- 103. Greenberg GR, Marliss EB, Anderson GH, Langer B, Spence W, et al. 1976. Protein-sparing therapy in postoperative patients. Effects of added hypocaloric glucose or lipid. New Engl. J. Med. 294:1411-16
- 104. Gregerman RK, Solomons N. 1967. Acceleration of thyroxine and triiodothyronine turnover during bacterial pulmonary infections and fever: implications for the functional state of the thyroid during stress and senescence. J. Clin. Endocrinol. Metab. 27:93-105
- Hagen GA, Elliott WJ. 1973. Transport of thyroid hormones in serum and cerebrospinal fluid. J. Clin. Endocrinol. Metab. 37:415-22
- 106. Hall-Angerås M, Angerås U, Zamir O, Hasselgren PO, Fisher JE. 1990. Interaction between corticosterone and tumor necrosis factor stimulated protein breakdown in rat skeletal muscle, similar to sepsis. Surgery 108:460-66
- 107. Hammond GL, Smith CL, Goping I, Underhill D, Harvey M, et al. 1987. Primary structure of human corticosteroid binding globulin, deduced from hepatic and pulmonary cDNAs, exhibits homology with serine protease inhibitors. Proc. Natl. Acad. Sci. USA 84: 5153-57
- Hammond GL, Smith CL, Paterson N, Sibbald WJ. 1990. A role for corticosteroid-binding globulin in delivery of cortisol to activated neutrophils. J. Clin. Endocrinol. Metab. 71:34-39
- Hammond GL, Smith CL, Underhill C, Nguyen VT. 1991. Interaction between corticosteroid binding globulin and activated leukocytes in vitro. Biochem. Biophys. Res. Commun. 172:172-77
- Haq RU, Chytil F. 1988. Early effects of retinol and retinoic acid on protein synthesis in retinol deficient rat testes. Biochem. Biophys. Res. Commun. 151: 53-60
- Harding J, Skare J, Shinner M. 1991. A second transthyretin mutation at position 33 (LewPhe) associated with familial amyloidotic polyneuropathy. *Biochim. Biophys. Acta* 1097:183–86
- 112. Harland WA, Horton PW, Strang R, Fitzegerald B, Richards J, Holloway K. 1974. Release of thyroxine from the liver during anesthesia and surgery. Br. J. Anaesth. 46:818-20
- Harrison HH, Gordon ED, Nichols WC, Benson MD. 1991. Biochemical and

- clinical characterization of prealbumin-Chicago: an apparently benign variant of serum prealbumin (transthyretin) discovered with high-resolution two-dimensional electrophoresis. Am. J. Med. Genet. 39:442-52
- Harvey RF. 1971. Serum-thyroxine and thyroxine-binding globulin in seriously ill patients. *Lancet* 1:208-12
- Hasselgren PO, Pedersen P, Sax HC, Warner BW, Fisher JE. 1988. Current concepts of protein turnover and amino acid transport in liver and skeletal muscle during sepsis. Arch. Surg. 123:992– 99
- Herbert J, Wilcox JN, Pham KT, Fremeau R, Zaviani M, et al. 1986. Transthyretin: a choroid plexus-specific transport protein in human brain. Neurology 36:900-11
- Herrmann FR, Safran C, Levkoff SE, Minaker KL. 1992. Serum albumin level on admission as a predictor of death, length of stay and readmission. Arch. Int. Med. 152:125-30
- Holmgren G, Haetiner E, Nordenson I, Sandgren O, Steen L, Lundgren E 1988. Homozygosity for the transthyretinmet 30-gene in two Swedish sibs with familial amyloidotic polyneuropathy. Clin. Genet. 34:333-38
- Homgren G, Ericzon BG, Groth CG, Steen L, Suhr O, et al. 1993. Clinical improvement and amyloid regression after liver transplantation in hereditary transthyretin amyloidosis. *Lancet* 341: 1113-16
- Hoover HC Jr, Ryan JA, Anderson EJ, Fisher JE. 1980. Nutritional benefits of invnediate postoperative jejunal feeding of an elemental diet. Am. J. Surg. 139: 153-59
- Ii S, Minnerath S, Ii K, Dyck PJ, Sommer S. 1991. Two-tiered DNA-based diagnosis of transthyretin amyloidosis reveals two novel point mutations. Neurology 41:893-98
- Inada M, Sterling K. 1967. Thyroxine transport in thyrotoxicosis and hypothyroidism. J. Clin. Invest. 46:1442-50
- Ingbar SH. 1958. Prealbumin: a thyroxine-binding protein of human plasma. Endocrinology 63:256-59
- Ingbar SH, Freinkel N. 1960. Regulation of the peripheral metabolism of the thyroid hormones. Recent Progr. Horm. Res. 16:353-403
- Ingenbleek Y, De Visscher M, De Nayer P. 1972. Measurement of prealbumin as index of protein-calorie malnutrition. Lancet 2:106-9
- Ingenbleek Y, De Nayer P, De Visscher M. 1974. Thyroxine-binding globulin in

- infant protein-calorie malnutrition. J. Clin. Endocrinol. Metab. 39:178-80
- 127. Ingenbleek Y, Van den Schrieck HG, De Nayer P, De Visscher M. 1975. Albumin, transferrin and the thyroxinebinding prealbumin/retinol-binding protein (TBPA-RBP) complex in assessment of malnutrition. Clin. Chim. Acta 63:61-67
- 128. Ingenbleek Y, De Nayer P, De Visscher M. 1975. Discrepancy between the measurement of thyroxine-binding prealbumin plasma level and binding capacity in protein-calorie malnutrition. Eur. J. Clin. Invest. 5:187-90
- 129. Ingenbleek Y. 1977. La malnutrition protéino-calorique chez l'enfant en bas âge. Répercussions sur la fonction thyroidienne et les protéines vectrices du sérum. PhD thesis, pp. 1-212. Univ. Louvain. Leuven: Acco
- Ingenbleek Y, De Visscher M. 1979. Hormonal and nutritional factors: critical conditions for endemic goiter epidemiology? Metabolism 28:9-19
- Ingenbleek Y. 1980. Thyroid function in nonthyroid illnesses. In *The Thyroid Gland*, ed. M De Visscher, pp. 499-527. New York: Raven
- Ingenbleek Y, Malvaux P. 1980. Peripheral turnover of thyroxine and related parameters in protein-calorie malnutrition. Am. J. Clin. Nutr. 33:609-16
- 133. Ingenbleek Y. 1982. Usefulness of prealbumin as nutritional indicator. In Marker Proteins in Inflammation, ed. R Allen, J Bienvenu, P Laurent, R Suskind, 1:405-14. Berlin: Walter de Gruyter
- Ingenbleek Y, Carpentier YA. 1985. A prognostic inflammatory and nutritional index scoring critically ill patients. *Int.* J. Vitam. Nutr. Res. 55:91-101
- Isakov N. 1988. Regulation of T-cell derived protein kinase C activity by vitamin A derivatives. Cell. Immunol. 115:288-98
- Ismadi SD, Olson JA. 1975. Vitamin A transport in human fetal blood. Am. J. Clin. Nutr. 8:967-72
- 137. Isshiki H, Akira S, Sugita T, Nishio Y, Hashimoto S, et al. 1991. Reciprocal expression of NF-II6 and C/EBP in hepatocytes: possible involvement of NF-II6 in acute phase protein gene expression. New Biol. 3:63-70
- Izumoto S, Younger D, Hays AP, Martone RL, Smith R, Herbert J. 1992. Familial amyloidotic polyneuropathy presenting with carpal tunnel syndrome and a new transthyretin mutation, Asparagine-70. Neurology 42:2094-102
- Jacobsen BB, Peitersen B, Andersen HJ,

- Hummer L. 1979. Serum concentrations of thyroxine-binding globulin, prealbumin and albumin in healthy full-term, small-for-gestational age and preterm newborn infants. Acta Paediatr. Scand. 68:49-55
- Jacobson DR, Reveille JD, Buxbaum JN. 1991. Frequency and genetic background of the position 122 (Val→lle) variant transthyretin gene in the Black population. Am. J. Hum. Genet. 49:192– 98
- Jacobson DR, McFarlin DE, Kane I, BuxbaumJN. 1992. Transthyretin Pro⁵⁵, a variant associated with early-onset, aggressive, diffuse amyloidosis with cardiac and neurologic involvement. Hum. Genet. 89:353-56
- Jacobsson B. 1989. A study of transthyretin gene expression in normal and neoplastic tissues. PhD thesis, pp. 1-48. Karolinska Inst., Stockholm
- Jeevanandam M, Horowitz G, Lowry SF, Brennan MF. 1984. Cancer cachexia and protein metabolism. Lancet 1:1423– 26
- 144. Jennings G, Bourgeois C, Elia M. 1992. The magnitude of the acute phase protein response is attenuated by protein deficiency in rats. J. Nutr. 122:1325-31
- 145. Jones LA, Skare JC, Harding J, Cohen A, Milunsky A, Skinner M. 1991. Proline at position 36: a new transthyretin mutation associated with familial amyloidotic polyneuropathy. Am. J. Hum. Genet. 48:979-82
- 146. Jones LA, Skare JC, Cohen AS, Harding J, Milunsky A, Skinner M. 1992. Familial amyloidotic polyneuropathy: a new transthyretin position 30 mutation (alanine for valine) in a family of German descent. Clin. Genet. 41:70-73
- Jörnvall H, Carlström A, Pettersson T, Jacobsson B, Persson M, Mutt V. 1981.
 Structural homologies between prealbumin, gastrointestinal prohormones and other proteins. Nature 291:261-63
- 148. Kabat EA, Moore D, Landow H. 1942. An electrophoretic study of the protein components in cerebrospinal fluid and their relationship to the serum proteins. J. Clin. Invest. 21:571-77
- Kanai M, Raz A, Goodman DS. 1968. Retinol-binding protein: the transport protein for vitamin A in human plasma. J. Clin. Invest. 47:2025-44
- Kanda Y, Goodman DS, Canfield R, Morgan F. 1974. The amino acid sequence of human plasma prealbumin. J. Biol. Chem. 249:6796–805
- Kasper H, Brodersen M, Schedel R. 1975. Concentration of vitamin A, retinol-binding protein and prealbumin in

- response to stress. Acta Hepato-Gastroenterol. 22:403-8
- 152. Katoh M, Kanai M, Kameko M, Ohno S, Fujii Y, Nagata T. 1982. Localization of retinol-binding protein and prealbumin in the human kidney with an unlabeled immunohistochemical method. Acta Histochem. Cytochem. 15: 68-75
- 153. Kelleher PC, Phinney SD, Sims E, Bogardus C, Horton E, et al. 1983. Effects of carbohydrate-containing and carbohydrate-restricted hypocaloric and eucaloric diets on serum concentrations of retinol-binding protein, thyroxine-binding prealburnin and transferrin. Metabolism 32:95-101
- Kiorpes TC, Anderson RS, Wolf G. 1981. Effect of vitamin A deficiency on glycosylation of rat serum α1-macroglobulin. J. Nutr. 111:2059-68
- Kirven MJ, Wolf G. 1991. Synthesis and glycosylation of fibronectin in hepatocytes of vitamin A-deficient rats. Mol. Cell. Biochem. 101:101-14
- 156. Kitagawa M, Ohba Y, Suzuki Y. 1987. T3 action and chromatin structure. In Hormonal Regulation of Gene Expression, ed. Cent. Acad. Publ., 24:155-68. Tokyo: Gunma Univ.
- Klasing KC. 1988. Nutritional aspects of leukocytic cytolaines. J. Nutr. 118: 1436–46
- 158. Kobayashi M, Horiuchi R, Hachisu T, Takikawa H. 1988. Dualistic effects of thyroid hormone on a human hepatoma cell line: inhibition of thyroxine-binding globulin synthesis and stimulation of α1-acid glycoprotein synthesis. Endocrinology 123:631-40
- Krust A, Kastner PH, Petkovich M, Zelent A, Chambon P. 1989. A third human retinoic acid receptor, hRAR-γ. Proc. Natl. Acad. Sci. USA 96:5310-14
- Kudlackova M, Andel M, Hajkova M, Novakova J. 1990. Acute phase proteins and prognostic inflammatory and nutritional index (PINI) in moderately burned children aged up to 3 years. Burns 16: 53-56
- Kushner I. 1992. The phenomenon of the acute phase response. Ann. N. Y. Acad. Sci. 389:39-49
- Larsen PD, Delallo L. 1989. Cerebrospinal fluid transthyretin in the neonate and blood-cerebrospinal fluid barrier permeability. Ann. Neurol. 25:629-30
- 163. Larsson M, Pettersson T, Carlström A. 1985. Thyroid hormone binding in serum of 15 vertebrate species. Isolation of thyroxine-binding globulin and prealbumin analogs. Gen. Comp. Endocrinol. 58:360-75

- Laurell CB. 1972. Electroimmunoassay. Scand. J. Clin. Lab. Invest. 29(Suppl. 124):21-37
- Le Moullac B, Gouache P, Bleiberg-Daniel F. 1992. Regulation of hepatic transthyretin messenger RNA levels during moderate protein and food restriction in rats. J. Nutr. 122:864-70
- 166. Leonard JL, Siegrist-Kaiser CA, Zuckerman CJ. 1990. Regulation of type II iodothyronine 5'-deiodase by thyroid hormone. J. Biol. Chem. 265:940-46
- Lohnes D, Dierich A, Ghyselinck N, Kastner P, Lampron C, et al. 1992. Retinoid receptors and binding proteins. J. Cell Sci. S16:69-76
- Long C. 1977. Energy balance and carbohydrate metabolism in infection and sepsis. Am. J. Clin. Nutr. 30:1301-10
- Macintyre SS, Kushner I, Samols D. 1985. Secretion of C-reactive protein becomes more efficient during the course of the acute phase response. J. Biol. Chem. 260:4169-73
- Makover A, Moriwalai H, Ramakrishnan R, Saraiva MJM, Blaner WS, Goodman DS. 1988. Plasma transthyretin: tissue sites of degradation and turnover in the rat. J. Biol. Chem. 263:8598-603
- 171. Makover A, Soprano DR, Wyatt ML, Goodman DS. 1989. An in situ hybridization study of the localization of retinol-binding protein and transthyretin messenger RNAs during fetal development in the rat. Differentiation 40: 17-25
- Martone RL, Herbert J, Dwork A, Schon EA. 1988. Transthyretin is synthesized in the mammalian eye. Biochem. Biophys. Res. Commun. 151:905-12
- Martone RL, Schon EA, Goodman DS, Soprano D, Herbert J. 1988. Retinolbinding protein is synthesized in the mammalian eye. Biochem. Biophys. Res. Commun. 157:1078-94
- 174. Mendel CM, Weisiger RA, Jones A, Cavalieri RR. 1987. Thyroid hormone binding proteins in plasma facilitate uniform distribution of thyroxine within tissues: a perfused rat liver study. Endocrinology 120:1742-49
- Mendel CM, Cavalieri RR, Weisiger RA. 1988. Uptake of thyroxine by the perfused rat liver: implications for the free hormone hypothesis. Am. J. Physiol. 255:El10-19
- 176. Mendel CM, Weisiger RA, Cavalieri RR. 1988. Uptake of 3,5,3'-triiodothyronine by the perfused rat liver: return to the free hormone hypothesis. Endocrinology 123:1817-24
- 177. Mendel CM, Kuhn RW, Weisiger RA, Cavalieri RR, Siiteri PK, et al. 1989. Uptake of cortisol by the perfused rat

- liver: validity of the free hormone hypothesis applied to cortisol. *Endocrinology* 124:468-76
- Mendel CM. 1989. The free hormone hypothesis: a physiologically based mathematical model. Endocr. Rev. 10: 232-74
- Mickell JJ. 1982. Urea nitrogen excretion in critically ill children. *Pediatrics* 70:949-55
- Miholic J, Hudec M, Müller MM, Domanig E, Wolner E. 1986. Early prediction of deep sternal wound infection after heart operations by alpha-1 acid glycoprotein and C-reactive protein measurements. Ann. Thorac. Surg. 42:429-33
- Miller MJS, Sadowska-Krowicka H, Chotinaruemol S, Kakis JL, Clark DA. 1993. Amelioration of chronic ileitis by nitric oxide synthase inhibition. J. Pharmacol Exp. Ther. 264:11-16
- macol. Exp. Ther. 264:11-16
 182. Mita S, Macda S, Shimada K, Araki S.
 1984. Cloning and sequence analysis of cDNA for human prealbumin. Biochem.
 Biophys. Res. Commun. 124:558-64
- 183. Mohanram M, Bamji MS. 1979. Serum vitamin A and retinol binding protein in malnourished women treated with oral contraceptives: effects of estrogen dose and duration of treatment. Am. J. Obstet. Gynecol. 135:470-72
- 184. Mohler JL, Michael KA, Freedman AM, Griffen W, McRoberts JW. 1985. The serum and urinary cortisol response to operative trauma. Surgery 161:445-49
- Moscowitz SR, Pereira G, Spitzer A, Heaf L, Amsel J, Watkins JB. 1983.
 Prealbumin as a biochemical marker of nutritional adequacy in premature infants. J. Pediatr. 102:749-53
- 186. Moses AC, Rosen HN, Moller D, Tsusaki S, Haddow J, et al. 1990. A point mutation in transthyretin increases affinity for thyroxine and produces euthyroid hyperthyroxinemia. J. Clin. Invest. 86:2025-33
- Moshage HJ, Janssen JA, Franssen J, Hafkenscheid J, Yap SM. 1987. Studies of the molecular mechanism of decreased liver synthesis of albumin in inflammation. J. Clin. Invest. 79:1635– 41
- Munro HN. 1951. Carbohydrate and fat as factors in protein utilization. *Physiol. Rev.* 32:449–88
- Munro HN. 1970. A general survey of mechanisms regulating protein metabolism in mammals. In Mammalian Protein Metabolism, ed. HN Munro, 4:299-321. New York: Academic
- Murakami T, Ohnishi S, Nishiguchi S, Maeda S, Araki S, Shimada K. 1988.

- Acute-phase response of mRNAs for serum amyloid P component, C-reactive protein and prealbumin (transthyretin) in mouse liver. Biochem. Biophys. Res. Commun. 155:554-60
- Murakami T, Maeda S, Yi S, Ikegawa S, Kawashima E, et al. 1992. A novel transthyretin mutation associated with familial amyloidotic polyneuropathy. Biochem. Biophys. Res. Commun. 182: 520-26
- Murray MJ, Marsh HM, Wochos DN, Moxness KE, Offord KP, Callaway CW. 1988. Nutritional assessment of intensive-care unit patients. Mayo Clin. Proc. 63:1106-15
- Murty CN, Sidransky H. 1972. The effect of tryptophan on messenger RNA of livers of fasted mice. Biochim. Biophys. Acta 262:328-35
- 194. Nakazato M, Kangawa K, Minamino N, Tawara S, Matsuo H, Araki S. 1984. Revised analysis of amino acid replacement in a prealbumin variant (SKO-III) associated with familial amyloidotic polyneuropathy of Jewish origin. Biochem. Biophys. Res. Commun. 123: 921-28
- 195. Nakazato M, Ikeda S, Shiomi K, Matsukura S, Yoshida K, et al 1992. Identification of a novel variant (Val3O->Leu) associated with familial amyloidotic polyneuropathy. FEBS Lett. 306:206-8
- Navab M, Mallia AK, Kanda Y, Goodman DS. 1977. Rat plasma prealbumin: isolation and partial characterization. J. Biol. Chem. 252:5100-6
- Navab M, Smith JE, Goodman DS. 1977. Rat plasma prealbumin. Metabolic studies on effects of vitamin A status and on tissue distribution. J. Biol. Chem. 252:5107-14
- Newcomer M, Jones TA, Aqvist J, Sundelin J, Eriksson U, et al. 1984. The three-dimensional structure of retinolbinding protein. EMBO J. 3:1451-54
- 199. Nichols WC, Liepnieks J, McKusick V, Benson MD. 1989. Direct sequencing of the gene for Maryland/German familial amyloidotic polyneuropathy type II and genotyping by allele-specific enzymatic amplification. Genomics 5:535-40
- Nichols WC, Padilla LM, Benson MD. 1989. Prenatal detection of a gene for hereditary amyloidosis. Am. J. Med. Genet. 34:520-24
- Nilsson SF, Rask L, Peterson PA. 1971.
 Evidence for multiple thyroxine binding sites in human prealbumin. J. Biol. Chem. 246:6098-105
- 202. Nishi H, Kimura A, Harada H, Hayashi

- Y, Nakamura M, Sasazuki T. 1992. Novel variant transthyretin gene (Ser50 to Ile) in familial cardiac amyloidosis. Biochem. Biophys. Res. Commun. 187: 460-66
- Nordlie M, Sletten K, Husby G, Ranlov PJ. 1988. A new prealburnin variant in familial cardiomyopathy of Danish origin. Scand. J. Immunol. 27:119-22
- Noy N, Xu ZJ. 1990. Interactions of retinol with binding proteins: implications for the mechanism of uptake by cells. Biochemistry 29:3878-83
- Nunez EA. 1989. The erb-A family receptors for thyroid hormones, steroids, vitamin D and retinoic acid: characteristics and modulation. Curr. Opin. Cell. Biol. 1:177-85
- Ong DE. 1985. Vitamin A--binding proteins. Nutr. Rev. 43:225-32
- Oppenheimer JH, Martinez M, Bernstein G. 1966. Determination of the maximal binding capacity and protein concentration of thyroxine-binding prealbumin in human serum. J. Lab. Clin. Med. 67: 500-9
- Oppenheimer JH, Werner S. 1966. Effect of prednisone on thyroxine-binding proteins. J. Clin. Endocrinol. 26:715-17
- Oppenheimer JH, Schwartz HL, Koerner D, Surks MI. 1974. Limited binding capacity sites for L-triiodothyronine in rat liver nuclei. Nuclear-cytoplasmic interrelation, binding constants, and cross-reactivity with L-thyroxine. J. Clin. Invest. 53:768-77
- Oppenheimer JH, Schwartz HL, Mariash CN, Kinlaw W, Wong N, Freake HC. 1987. Advances in our understanding of thyroid hormone action at the cellular level. Endocr. Rev. 8:288-308
- Osborne TB, Mendel LB. 1914. Aminoacids in nutrition and growth. J. Biol. Chem. 17:325-49
- Peeples JM, Carlson SE, Werkman SH, Cooke RJ. 1991. Vitamin A status of preterm infants during infancy. Am. J. Clin. Nutr. 53:1455-59
- Pemberton PA, Stein PE, Pepys M, Potter J, Carrell R. 1988. Hormone binding globulins undergo serpin conformational change in inflammation. *Nature* 336: 257-58
- 214. Pencharz PB, Motil KJ, Parsons H, Duffy BJ. 1980. The effect of an energy-restricted diet on the protein metabolism of obese adolescents: nitrogenbalance and whole-body nitrogen turnover. Clin. Sci. 59:13-18
- Peters T. 1975. Serum albumin. In The Plasma Proteins: Structure, Function and Genetic Control, ed. FW Putnam, 1:133-81. New York: Academic

- Peterson PA. 1971. Demonstration in serum of two physiological forms of the human retinol binding protein. Eur. J. Clin. Invest. 1:437-44
- Peterson PA, Nilsson S, Ostberg L, Rask L, Vahlquist A. 1974. Aspects of the metabolism of retinol-binding protein and retinol. Vitam. Horm. 32:181-214
- Pettersson T, Carlström A, Jörnvall H. 1987. Different types of microheterogeneity of human thyroxine-binding prealbumin. Biochemistry 26:4572–83
- Pozzetto B, Odelin MF, Bienvenu J, Defayolle M, Aymard M. 1993. Is there a relationship between malnutrition, inflammation, and post-vaccinal antibody response to influenza viruses in the elderly? J. Med. Virol. 41:39-43
- Pras M, Franklin EC, Prelli F, Frangione B. 1981. A variant of prealbumin from amyloid fibril in familial amyloidotic polyneuropathy of Jewish origin. J. Exp. Med. 154:989-93
- Prescott RW, Yeo PP, Watson MJ, Johnston I, Ratcliffe J, Evered D. 1979. Total and free thyroid hormone concentrations after elective surgery. J. Clin. Pathol. 32:321-24
- Pressac M, Vignoli L, Aymard P, Ingenbleek Y. 1990. Usefulness of a prognostic inflammatory and nutritional index in pediatric clinical practice. Clin. Chim. Acta 188:129-36
- 223. Pugeat M, Bonneton A, Perrot D, Rocle-Nicolas B, Lejeune H, et al. 1989. Decreased immunoreactivity and binding activity of corticosteroid-binding globulin in serum in septic shock. Clin. Chem. 35:1675-79
- Purdy RH, Woeber KA, Holloway MT, Ingbar SH. 1965. Preparation of crystalline thyroxine-binding prealbumin from human plasma. Biochemistry 4: 1888-95
- 225. Ramsden DB, Prince HP, Burr W, Bradwell A, Black E, Hoffenberg R. 1978. The inter-relationship of thyroid hormones, vitamin A and their binding proteins following acute stress. Clin. Endocrinol. 8:109-22
- Rask L, Anundi H, Peterson PA. 1979.
 The primary structure of the human retinol-binding protein. FEBS Lett. 104: 55-58
- Rask L, Anundi H, Bohme J, Eriksson U, Frederiksson A, et al. 1980. The retinol-binding protein. Scand. J. Clin. Lab. Invest. 40:45-61
- Recant L, Riggs DS. 1952. Thyroid function in nephrosis. J. Clin. Invest. 31:789-97
- 229. Refetoff S, Dwulet FE, Benson MD. 1986. Reduced affinity for thyroxine

- in two of three structural thyroxinebinding prealbumin variants associated with familial amyloidotic polyneuropathy. J. Clin. Endocrinol. Metab. 63: 1432-37
- Rickard KA, Grosfeld JL, Kirksey A, Ballantine T, Baehner R. 1979. Reversal of protein-energy malnutrition in children during treatment of advanced neoplastic disease. Ann. Surg. 190:771-81
- Robbins J. 1956. Reverse-flow electrophoresis: a method for determining the thyroxine-binding capacity of serum protein. Arch. Biochem. Biophys. 63: 461-69
- Robbins J, Rall JE. 1957. The interaction of thyroid hormones and protein in biological fluids. Recent Progr. Horm. Res. 13:161-202
- Robbins J, Bartalena L. 1986. Plasma transport of thyroid hormones. In Thyroid Hormone Metabolism, Basic and Clinical Endocrinology, ed. G Hennemann, pp. 3-38. New York: Dekker
- mann, pp. 3-38. New York: Dekker 234. Roh MS, Drazenovich KA, Barbose JJ, Dinarello CA, Cobb CP. 1987. Direct stimulation of the adrenal cortex by interleukin-1. Surgery 102:140-46
- Rosner W. 1990. The functions of corticosteroid-binding globulin and sex hormone-binding globulin: recent advances. Endocr. Rev. 11:80-91
- Rossi R, Hirvonen T, Toivanen P. 1969.
 The concentration of prealbumin in human fetal and infant sera. Scand. J. Clin. Lab. Invest. 26:35-46
- Rothschild MA, Oratz M, Schreiber SS.
 1972. Albumin synthesis. New Engl. J. Med. 286:748-57, 816-21
- Rousseau GG. 1984. Control of genes expression by glucocorticoid hormones. Biochem. J. 224:1-12
- Rowlands BJ, Clark RG. 1978. Postoperative amino acid infusions: an appraisal. Br. J. Surg. 65:384-89
- praisal. Br. J. Surg. 65:384-89

 240. Saeki Y, Ueno S, Yorifuji S, Sugiyama Y, Ide Y, Matsuzawa Y. 1991. New mutant gene (transthyretin Arg-58) in cases with hereditary polyneuropathy detected by non-isotope method of single-strand conformation polymorphism analysis. Biochem. Biophys. Res. Commun. 180:380-85
- Sakurada T, Saito S, Inagaki K, Tayama S, Torikai T. 1967. Polyacrylamide gel electrophoresis study on the effect of estrogen on human thyroxine-binding prealbumin. Tohoku J. Exp. Med. 93: 839-62
- Saraiva MJ, Costa P, Birken S, Goodman DS. 1983. Presence of an abnormal transthyretin (prealbumin) in Portuguese

Trans. Assoc. Am. Physicians 96:261-70 243. Saraiva MJ, Almeida MR, Sherman W, Gawinowicz M, Costa P, et al. 1992. A

familial amyloidotic polyneuropathy.

- Gawinowicz M, Costa P, et al. 1992. A new transthyretin mutation associated with amyloid cardiomyopathy. Am. J. Hum. Genet. 50:1027-30
- Sasaki H, Yoshioka M, Takagi Y, Sakaki Y. 1985. Structure of the chromosomal gene for human serum prealbumin. Gene 37:191-97
- 245. Sasanow SR, Spitzer AR, Pereira GR, Heaf L, Watkins JB. 1986. Effect of gestational age upon prealbumin and retinol binding protein in preterm and term infants. J. Pediatr. Gastroenterol. Nutr. 5:111-15
- Saudek CD, Felig P. 1976. The metabolic events of starvation. Am. J. Med. 60:117-26
- 247. Savu L, Vranckx R, Maya M, Nunez EA. 1987. A thyroxine binding globulin (TBG)-like protein in the sera of developing and adult rats. Biochem. Biophys. Res. Commun. 148:1165-73
- Schlechte JA, Hamilton D. 1987. The effects of glucocorticoids on corticosteroid binding globulin. Clin. Endocrinol. 27:197–203
- Schreiber G, Aldred AR, Jaworowski A, Nilsson C, Achen M, Segal MB. 1990. Thyroxine transport from blood to brain via transthyretin synthesis in choroid plexus. Am. J. Physiol. 258: R338-45
- Schultze HE, Schönenberger M, Schwick G. 1956. Über ein präalbumin des menschlichen serums. Biochem. Z. 328:267-84
- 251. Seelig S, Liaw C, Towle HC, Oppenheimer JH. 1981. Thyroid hormone attenuates and augments hepatic gene expression at a pretranslational level. *Proc. Natl. Acad. Sci. USA* 78:4733-37
- 252. Sevaljevic L, Ivanovic-Matic S, Petrovic M, Glibetic M, Pantelic D, Poznanovic G. 1989. Regulation of plasma acute-phase protein and albumin levels in the liver of scalded rats. Biochem. J. 258:663-68
- Shenai JP, Chytil F, Jhaveri A, Stahlman MT. 1981. Plasma vitamin A and retinol-binding protein in premature and term neonates. J. Pediatr. 99:302-5
- Shenton A, Wells FE, Addison GM. 1983. Prealbumin as an indicator of marginal malnutrition in treated phenylketonuria: a preliminary report. J. Inherit. Metab. Dis. 6(Suppl. 2):109-10
- Sherry B, Jack RM, Weber A, Smith AL. 1988. Reference interval for prealbumin for children two to 36 months old. Clin. Chem. 34:1878-80

- Shetty PS, Watrasiewicz KE, Jung R, James WPT. 1979. Rapid turnover transport proteins: an index of subclinical protein-energy malnutrition. *Lancet* 2: 230-32
- Shiels BR, Northemann W, Gehring MR, Fey GF. 1987. Modified nuclear processing of α1-acid glycoprotein RNA during inflammation. J. Biol. Chem. 262: 12826-31
- 258. Shiomi K, Nakazato M, Matsukara S, Ohnishi A, Hatanaka H, et al. 1993. A basic transthyretin variant (Glu⁶¹→Lys) causes familial amyloidotic polyneuropathy: protein and DNA sequencing and PCR-induced mutation restriction analysis. B ochem. Biophys. Res. Commun. 194:1090-96
- Sibbald WJ, Short A, Cohen M, Wilson RF. 1977. Variations in adrenocortical responsiveness during severe bacterial infections. Ann. Surg. 186:29-33
- Sidransky H, Verney E, Sarma D. 1971.
 Effect of tryptophan on hepatic polyribosomes and protein synthesis in liver.
 Am. J. Clin. Nutr. 24:779-85
- Siiteri PK, Murai JT, Hammond GL, Nisker J, Raymoure W, Kuhn RW. 1982. The serum transport of steroid hormones. Recent Progr. Horm. Res. 38: 457-510
- 262. Skare JC, Milunsky J, Milunsky A, Skare I, Cohen A, Skinner M. 1991. A new transthyretin variant from a patient with familial amyloidotic polyneuropathy has asparagine substituted for histidine at position 90. Clin. Genet. 39:6-12
- Shillmann JJ, Rosenoer VM, Smith PC, Fang MS. 1976. Improved albumin synthesis in postoperative patients by amino acid infusions. New Engl. J. Med. 295: 1037-40
- Sleinner M, Harding J, Skare J, Jones L, Cohen A, et al. 1992. A new transthyretin mutation associated with amyloidotic vitreous opacities. Asparagine for isoleucine at position 84. Ophtalmology 99:503-8
- Sklan D, Shalit I, Lasebnik N, Spirer Z, Weisman Y. 1985. Retinol transport proteins and concentrations in human amniotic fluid, placenta, and fetal and maternal sera. Br. J. Nutr. 54:577-83
- Sklan D, Ross C. 1986. Synthesis of retinol-bin ing protein and transthyretin in yolk sac and fetus in the rat. J. Nutr. 117:436-42
- Skrede S, Blomhoff JP, Elgjo K, Gjone E. 1975. Serum proteins in diseases of the liver. Scand. J. Clin. Lab. Invest. 35:399-406
- Smith FR, Goodman DS. 1971. Effects of diseases of the liver, thyroid and

- kidneys on the transport of vitamin A in human plasma. J. Clin. Invest. 50: 2426-36
- Smith FR, Suskind RM, Thanangkul O, Leitzman C, Goodman DS, Olson RE. 1975. Plasma vitamin A, retinol-binding protein, and pre-albumin concentrations in Protein-calorie malnutrition. III. Response to varying dietary treatments. Am. J. Clin. Nutr. 28:732-38
- Smith JE, Goodman DS. 1979. Retinolbinding protein and the regulation of vitamin A transport. Fed. Proc. 38: 2504-9
- Socolow EL, Woeber KA, Purdy RH, Holloway MT, Ingbar SH. 1965. Preparation of I-131-labeled human serum prealbumin and its metabolism in normal and sick patients. J. Clin. Invest. 44: 1600-9
- 272. Soprano DR, Herbert J, Soprano K, Schon EA, Goodman DS. 1985. Demonstration of transthyretin mRNA in brain and other extrahepatic tissues in the rat. J. Biol. Chem. 260:11793-98
- Soprano DR, Pickett CB, Smith JE, Goodman DS. 1982. Biosynthesis of plasma retinol binding protein in liver as a larger molecular weight precursor. J. Biol. Chem. 256:8256-58
- Squef R, Martinez M, Oppenheimer JH. 1963. Use of thyroxine-displacing drugs in identifying serum thyroxine-binding proteins separated by starch gel electrophoresis. Proc. Soc. Exp. Biol. 118:837– 80
- Stabilini R, Vergani C, Agostini A, Pugno Vanoni Agostini R. 1968. Influence of age and sex on prealbumin levels. Clin. Chim. Acta 20:358-59
- Staehelin T, Verney E, Sidransky H. 1967. The influence of nutritional change on polyribosomes of the liver. Biochim. Biophys. Acta 145:105-19
- 277. Stauder AJ, Dickson PW, Aldred AR, Schreiber G, Mendelsohn F, Hudson P. 1986. Synthesis of transthyretin (prealbumin) mRNA in choroid plexus epithelial cells, localized by in situ hybridization in the rat brain. J. Histochem. Cytochem. 34:949-52
- Sterling K, Tabachnick M. 1961. Paper electrophoretic demonstration of thyroxine-binding prealbumin fraction in serum. Endocrinology 68:1073-75
- Sterling KM Jr, Harris MJ, Mitchell J, Dipetrillo T, Delaney T, Cutroneo K. 1983. Dexamethasone decreases the amounts of type I procollagen mRNAs in vivo and in fibroblast cell cultures. J. Biol. Chem. 258:7644-47
- 280. Stoner HB. 1986. Metabolism after

- trauma and in sepsis. Circ. Shock 19:75-87
- Strahler JR, Rosenblum B, Hanash S. 1987. Identification and characterization of a human transthyretin variant. Biochem. Biophys. Res. Commun. 148: 471-77
- Stroupe SD, Harding GB, Forsthoefel M, Westphal U. 1978. Kinetic and equilibrium studies on steroid interaction with human corticosteroid-binding globulin. Biochemistry 17:177-83
- Sundelin J, Melhus H, Das S, Eriksson U, Lind P, et al. 1985. The primary structure of rabbit and rat prealbumin and a comparison with the tentiary structure of human prealbumin. J. Biol. Chem. 260:6481-87
- 284. Talwar KK, Sawhney RC, Rastogi GK. 1977. Serum levels of thyrotropin, thyroid hormones and their response to thyrotropin releasing hormone in infective febrile illnesses. J. Clin. Endocrinol. Metab. 44:398-403
- Tawara S, Nakazato M, Kangawa K, Matsuo H, Aralii S. 1983. Identification of amyloid prealbumin variant in familial amyloidotic polyneuropathy (Japanese type). Biochem. Biophys. Res. Commun. 116:880-88
- Tracey KJ, Vlassara H, Cerami A. 1989.
 Cachectin/tumour necrosis factor. Lancet 1:1122-26
- Travis J, Guzdek A, Potempa J, Watorek W. 1990. Serpins: structure and mechanism of action. Biol. Chem. Hoppe-Seyler 371(S):3-11
- Trechsel U, Evequoz V, Fleisch H. 1985. Stimulation of interleukin 1 and 3 production by retinoic acid in vitro. Biochem. J. 230:339-44
- Tsujimoto M, Okamura N, Adachi H. 1988. Dexamethasone inhibits the cytotoxic activity of tumor necrosis factor. Biochem. Biophys. Res. Commun. 153: 109-15
- Tsuzuki T, Mita S, Maeda S, Araki S, Shimada K. 1985. Structure of the human prealbumin gene. J. Biol. Chem. 260:12224-27
- Uemichi T, Murrell JR, Zeldenrust S, Benson MD. 1992. A new mutant transthyretin (Arg-10) associated with familial polyneuropathy. J. Med. Genet. 29: 888-91
- 292. Ueno S, Uemichi T, Yorifuji S, Tarui S. 1990. A novel variant of transthyretin (Tyr¹¹⁴ to Cys) deduced from the nucleotide sequences of gene fragments from familial amyloidostic polyneuropathy in Japanese sibling cases. Biochem. Biophys. Res. Commun. 169: 143-47

- 293. Ueno S, Uemichi T, Takahashi N, Soga F, Yorifuji S, Tarui S. 1990. Two novel variants of transthyretin identified in Japanese cases with familial amyloidotic polyneuropathy: transthyretin (Glu⁴² to Gly) and transthyretin (Ser⁵⁰ to Arg). Biochem. Biophys. Res. Commun. 169: 1117-21
- 294. Vahlquist A, Rask L, Peterson PA, Berg T. 1975. The concentrations of retinol binding protein, prealbumin, and transferrin in the sera of newly delivered mothers and children of various ages. Scand. J. Clin. Lab. Invest. 35:569-75
- Vahlquist A, Sjölund K, Norden A, Peterson PA, Stigmar G, Johansson B. 1978. Plasma vitamin A and visual dark adaptation in diseases of the intestine and liver. Scand. J. Clin. Lab. Invest. 88:301-8
- van Jaarsveld P, Edelhoch H, Goodman DS, Robbins J. 1973. The interaction of human plasma retinol-binding protein with prealbumin. J. Biol. Chem. 248: 4698-705
- Vanlandingham S, Spiekerman A, Newmark S. 1982. Prealbumin: a parameter of visceral protein levels during albumin infusion. J. Parenter. Enter. Nutr. 6: 230-31
- Vannice JL, Ringold GM, McLean JW, Taylor JM. 1983. Induction of the acute phase reactant, α1-acid glycoprotein, by glucocorticoids in rat hepatoma cells. DNA 2:205-12
- Vehe KL, Brown RO, Kuhl DA, Boucher BA, Luther RW, Kudsk KA. 1991.
 The prognostic inflammatory and nutritional index in traumatized patients receiving enteral nutrition support. J. Am. Coll. Nutr. 10:355-63
- Vignati L, Finley RJ, Hagg S, Aoki TT. 1978. Protein conservation during prolonged fast: a function of triiodothyronine levels. Trans. Assoc. Am. Physicians 91:169-79
- Wade CE, Lindberg JS, Cockrell J, Lamiell J, et al. 1988. Upon admission adrenal steroidogenesis is adapted to the degree of illness in intensive case unit patients. J. Clin. Endocrinol. Metab. 67: 223-27
- Wade S, Parent G, Bleiberg-Daniel F, Maire B, Fall M, et al. 1988. Thymulin (Zn-FTS) activity in protein-energy malnutrition: new evidence for interaction between malnutrition and infection on thymic function. Am. J. Clin. Nutr. 47: 305-11
- 303. Wade S, Bleiberg-Daniel F, Le Moullac B, Iyakaremye D, Biou D, et al. 1988. Value of serum transthyretin measurements in the assessment of marginal

protein-energy malnutrition in rats. J. Nutr. 118:1002-10

304. Wade S, Bleiberg-Daniel F, Le Moullac B. 1988. Rat transthyretin: the effects of acute short-term food deprivation and refeeding on the serum and cerebrospinal fluid concentration and on the hepatic mRNA level. J. Nutr. 118: 199– 205

- Wakasugi S, Maeda S, Shimada K, Nakashima M, Migita S. 1985. Structural comparisons between mouse and human prealbumin. J. Biochem. 98: 1707-14
- Wallace MR, Naylor S, Kluve-Beckerman B, Long GL, McDonald L, et al. 1985. Localization of the human prealbumin gene to chromosome 18. Biochem. Biophys. Res. Commun. 129: 753-58
- Wallace MR, Dwulet FE, Conneally P, Benson MD. 1986. Biochemical and molecular genetic characterization of a new variant prealbumin associated with hereditary amyloidosis. J. Clin. Invest. 78: 6-12
- Wallace MR, Dwulet FE, Williams E, Conneally P, Benson MD. 1988. Identification of a new hereditary amyloidosis prealbumin variant, Tyr-77, and detection of the gene by DNA analysis. J. Clin. Invest. 81:189-93
- Warren RS, Donner DB, Starnes HF Jr, Brennan M. 1987. Modulation of endogenous hormone action by recombinant human tumor necrosis factor. Proc. Natl. Acad. Sci. USA 84:8619-22
- Wartofsky L, Martin D, Earll JM. 1972. Alterations in thyroid iodine release and the peripheral metabolism of thyroxine during acute falciparum malaria in man. J. Clin. Invest. 51:2215-32
- Watson F, Dick M. 1980. Distribution and inheritance of low serum thyroxinebinding globulin levels in Australian Aborigenes. Med. J. Aust. 2:385-87
- Weiser JN, Do YS, Felmann D. 1979.
 Synthesis and secretion of corticosteroid-binding globulin by rat liver. J. Clin. Invest. 63:461-67
- Weisner B, Roethig HJ. 1983. The concentration of prealbumin in cerebrospinal fluid, indicator of CSF circulation disorders. Eur. Neurol. 22:96-105
- Westermark P, Sletten K, Johansson B, Cornwell GG III. 1990. Fibril in senile systemic amyloidosis is derived from normal transthyretin. Proc. Natl. Acad. Sci. USA 87:2843-45
- Wilmore DW. 1983. Alterations in protein, carbohydrate, and fat metabolism in injured and septic patients. J. Am. Coll. Nutr. 2:3-13

- Wise L, Margraf HW, Ballinger WF. 1972. Adrenal cortical function in severe burns. Arch. Surg. 105:213-20
- Woeber KA, Doherty GF, Ingbar SH. 1972. Stimulation by phagocytosis of the deiodination of L-thyroxine in human leukocytes. Science 176: 1039– 41
- Yamamoto M. 1991. Retinoids in the host defense system. World Rev. Nutr. Diet. 64:58-84
- Yang RD, Moldawer LL, Sakamoto A, Keenan R, Matthews D, et al. 1983. Leucocyte endogenous mediator alters protein dynamics in rats. Metabolism 32: 654-60
- 320. Yap SH, Moshage HJ, Janssen JA, Franssen JH. 1986. The role of glucose in regulation of liver protein synthesis. In Nutritional Diseases: Research Directions in Comparative Pathobiology, pp. 91-101. New York: Liss
- Young GA, Chem C, Collins JP, Hill GL. 1979. Plasma proteins in patients receiving amino acids or intravenous hyperalimentation after major surgery. Am. J. Clin. Nutr. 32:1192-98
- 322. Young VR, Bier DM, Pellett PL. 1989. A theoretical basis for increasing current estimates of the amino acid requirements in adult man, with experimental support. Am. J. Clin. Nutr. 50: 80-92
- Young VR, Yu Y-M, Fukagawa NK. 1991. Protein and energy interactions throughout life. Metabolic basis and nutritional implications. Acta Paediatr. Scand. S373:5-24
- 324. Young VR, Yu Y-M, Krempf M. 1991. Protein and amino acid turnover using stables isotopes ¹⁵N, ¹³C and ²H as probes. In New Techniques in Nutritional Research, ed. RG Whitehead, A Prentice, pp. 17–72. San Diego: Academic
- 325. Young VR, Yu Y-M, Fukagawa NK. 1992. Energy and protein turnover. In Energy Metabolism: Tissue Determinants and Cellular Corollaries, ed. JM Kinney, HN Tucker, pp. 439-66. New York: Raven
- Zeldenrust S, Skinner M, Harding J, Skare J, Benson MD. 1993. A new transthyretin variant (His-69) associated with vitreous amyloid in an FAP family. J. Rheumatol. 20:188
- Zouaghi H, Savu L, Delorme J, Kleinknecht D, Nunez EA. 1983. Loss of serum transcortin in human shock associated with severe infection by candida albicans. Acta Endocrinol. 102: 277-83